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CURRENT OPINION

What pre-Columbian mummies could teach us about South American leishmaniases?

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One sentence summary: The evolutionary origin of visceral leishmaniasis in South America is discussed. Editor: Edmundo Calva

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ABSTRACT

A recent report on the taxonomic profile of the human gut microbiome in pre-Columbian mummies (Santiago-Rodriguez et al. 2016) gives for the first time evidence of the presence of *Leishmania* DNA (sequences similar to *Leishmania donovani* according to the authors) that can be reminiscent of visceral leishmaniasis during the pre-Columbian era. It is commonly assumed that *Leishmania infantum*, the etiological agent of American visceral leishmaniasis (AVL) was introduced into the New World by the Iberian conquest. This finding is really surprising and must be put into perspective with what is known from an AVL epidemiological and historical point of view. Beside *L. infantum*, there are other species that are occasionally reported to cause AVL in the New World. Among these, *L. colombiensis* is present in the region of pre-Columbian mummies studied. Other explanations for these findings include a more ancient introduction of a visceral species of *Leishmania* from the Old World or the existence of a yet unidentified endemic species causing visceral leishmaniasis in South America. Unfortunately, very few molecular data are known about this very long pre-Columbian period concerning the circulating species of *Leishmania* and their diversity in America.

Keywords: pre-Columbian mummies; pathogenic Leishmania species; American visceral leishmaniasis

ABBREVIATIONS

HIV: Human immunodeficiency virus DNA: Deoxyribonucleic Acid Mya: Million years ago Kya: Kilo years Santiago-Rodriguez et al. (2016) recently gave evidences of the presence of *Leishmania* DNA in the gut of two pre-Columbian mummified individuals from Cuzco. They use the complete genome of a species closely related to *Leishmania infantum* (*L. donovani*), as template, to assign DNA fragments from next

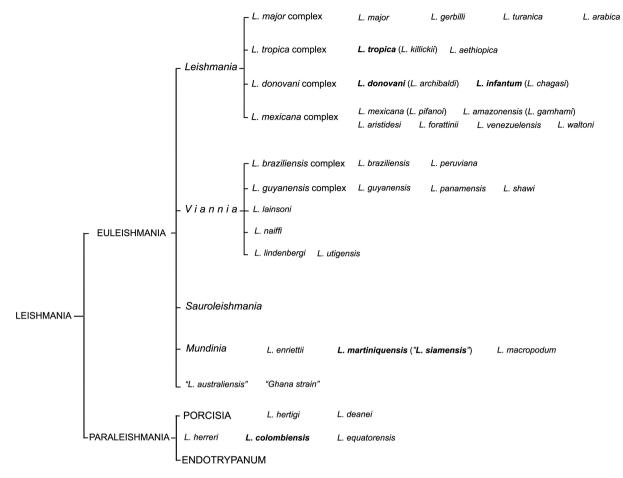


Figure 1. Updated classification of human pathogenic Leishmania species. Species known to cause visceral form of the disease are highlighted in bold.

generation sequencing (NGS) to *Leishmania*. They identified 19 DNA fragments of 50–434 bp matching with 16 contigs and gave no information on the reads (Santiago-Rodriguez *et al.* 2016). The circulation of parasites with visceralization capabilities, prior to the Iberian colonization, has never been mentioned before this report. In this viewpoint, we discuss the implication that this finding has on our current knowledge concerning the evolutionary history of the *Leishmania* genera and subgenera. We further address questions regarding the use of ancient microbial (*Leishmania*) DNA studies to follow the spread and prevalence of visceral leishmaniasis within human populations through time.

LEISHMANIA AND LEISHMANIASES

Leishmaniases are vector-borne diseases caused by obligate parasites (unicellular eukaryote) from the genus *Leishmania* (Trypanosomatida: Trypanosomatidae), which is subdivided into two major phylogenetic lineages, referred as the *Euleishmania* and *Paraleishmania* sections (Cupolillo et al. 2000). The section *Euleishmania* comprises five subgenera: *Leishmania sensu* stricto, Viannia, Sauroleishmania, Mundinia and the *Leishmania* australiensis-related species (see Akhoundi et al. 2016; Espinosa et al. 2016; Barratt et al. 2017). The subgenus Viannia is restricted to the Neotropics, while the subgenus *Leishmania sensu* stricto occurs in both the New and the Old World. The subgenus Sauroleishmania encompasses *Leishmania* species that are lizards infecting parasites (see Akhoundi et al. 2016) (see Fig. 1). The genus *Leishmania* includes 53 named species (without considering the synonyms) of which 29 are present in the Old World and 20 in the New World. Twenty-one species are pathogenic to humans.

The clinical manifestations of leishmaniases range from simple cutaneous lesions (CL) to diffuse cutaneous (DCL), mucocutaneous (MCL), mucosal (ML), post Kala-azar dermal (PKDL) and visceral (VL) or American visceral (AVL) forms.

The diseases are endemic in large areas of the tropics, subtropics and in the Mediterranean basin, globally spanning more than 98 countries and territories. There are about 350 million people at risk and some 12 million cases, with an estimated worldwide annual incidence of 0.7–1.2 million cases of CL and 0.2–0.4 million cases of VL (Alvar et al. 2012). Leishmania donovani in the Old World and L. infantum in both the Old and the New World are responsible for a large majority of visceral forms of the disease. Other cases of VL are caused occasionally by L. siamensis (Old World), L. martiniquensis (New World) and L. colombiensis (New World) (Cnudde et al. 1994; Akhoundi et al. 2016) (Fig. 2).

SOUTH AMERICAN LEISHMANIASES

In South America, all clinical forms of leishmaniasis are encountered. About 33% of CL cases occur in Brazil, but CL also have a great health impact on the entire subcontinent. More than 90% of VL cases occur in Brazil. In other countries (Alvar *et al.* 2012; Ready 2014), a very low endemicity is observed whereas

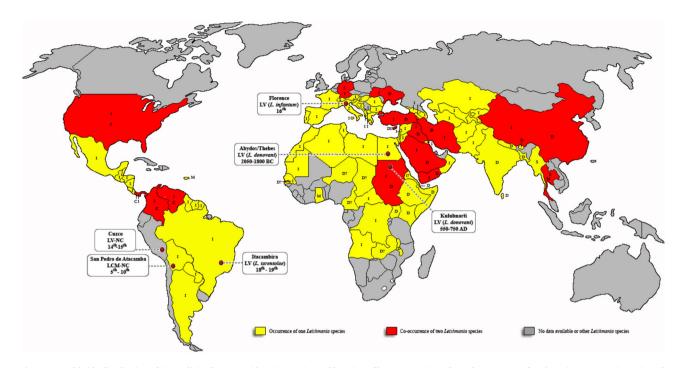


Figure 2. Worldwide distribution of visceralizing human Leishmania species and location of human remains where the presence of Leishmania DNA were investigated. NC: Identification of the Leishmania at the species level not sufficiently precise. I: L. infantum, D: L. donovani, C: L. colombiensis, S: L. siamensis, M, L. martiniquensis.

AVL is absent in Bolivia. In South America, the first descriptions of lesions reminiscent to those observed for current cutaneous forms are described as earlier as 1000 BC. The notification of a disease named 'valley sickness' or 'Andean sickness' or 'white leprosy' is mentioned during the 15th and 16th centuries AD that correspond to the Inca period. Proofs of a *Leishmania* infection are documented in a 6-year-old pre-Columbian female mummy (700–800 AD) (Gerszten, Allison and Maguire 2012). All these reports are likely to correspond to South American CL or MCL. The current cases of AVL diagnosed are all caused by *Leishmania* infantum initially named *L. chagasi*. Until now, AVL has never been documented in pre-Columbian individuals.

Three major hypotheses are proposed concerning the evolutionary history of the *Leishmania* genera and subgenera. Nevertheless, there are still missing elements from the puzzle to choose between the three evolutive scenari, which are as follows:

- The Palearctic hypothesis proposes the entrance of the Leishmania ancestor (evolved from lizard Leishmania) in the Nearctic across the Beringia land bridge during the Oligocene (33–23 Mya), spreading across the Northern continent until it reaches the Neotropics (Kerr, Merkelz and MacKinnon 2000) via the isthmus of Panama around 2.8 Mya (O'Dea et al. 2016).
- ii. The Neotropic hypothesis assumes that the genus *Leishma*nia has probably evolved in South America during the Paleocene or Eocene era (46–36 Mya). Then it has migrated via Beringia into Asia and dispersed towards India and Africa where major diversifications have occurred during the Miocene (24–14 Mya), giving rise to the various *Leishma*nia complexes (Croan, Morrison and Ellis 1997; Noyes et al. 2000; Lukes et al. 2007). *Leishmania infantum* has split from the early *L. donovani* lineage about 1 Mya ago (Kerr, Merkelz and MacKinnon 2000).

iii. The supercontinent hypothesis is based first on the division of *Leishmania* into two lineages on Gondwana (*Euleishmania* and *Paraleishmania*) and followed by the progressive separation of South America after the breakup of Gondwana during the late Cretaceous (100–90 Mya) (Harkins et al. 2016). A subsequent *Leishmania* migration back to the New World would be required during the mid-Miocene via the Nearctic and Beringia.

Regarding the origin of L. infantum in South America, it is believed that 500 years ago, L. infantum has crossed the Atlantic Ocean through Iberian colonization via infected dogs and reached Central America during the 16th century, causing AVL (Momen, Grimaldi and Deane 1987; Mauricio, Stothard and Miles 2000; Alvar et al. 2008; Kuhls et al. 2011; Leblois et al. 2011). As mentioned above, beside L. infantum, L. colombiensis (Paraleishmania section) causes VL in South America, although only few cases have been reported and documented (Kreutzer et al. 1991; Delgado et al. 1993; Rodriguez-Bonfante et al. 2003). In addition, L. martiniquensis, which belongs to the Mundinia section (Fig. 1), is the causative agent of VL in Martinique island (Cnudde et al. 1994). Unusual and rare clinical presentations, with a dissemination of parasites into internal organs, are well documented in some individuals infected by a Leishmania species that usually causes cutaneous forms. These are documented in the Old and New World (Barral et al. 1986, 1991; Barro-Traoré et al. 2008; Sarkari et al. 2016; Spotin and Parvizi 2016). Such unusual clinical presentation is frequently but not exclusively associated with the immune-suppression caused by human immunodeficiency virus co-infections for instance (Hernández et al. 1993, 1995; Ramos-Santos et al. 2000; Alvar et al. 2008; Zijlstra 2014). The dysfunction of the T-helper 1-mediated immune response for example is also likely to interfere with the progression, severity and clinical features of leishmaniases (Fishman 2011; Floral et al. 2014; Geiger et al. 2016).

AVL: WHAT PRE-COLUMBIAN MUMMIES TEACH US?

For the first time, Santiago-Rodriguez et al. (2016) gave evidences for the existence of Leishmania parasites with visceralization capabilities, prior to the Iberian colonization. Using as template the complete genome of Leishmania donovani, a species closely related to L. infantum, they assigned DNA fragments extracted from the gut of two pre-Columbian mummified individuals from Cuzco, as Leishmania. The post-Columbian origin of L. infantum in South America and the absence of its proven or suspected vectors (i.e. Lutzomyia almerioi, Lu. atunesi, Lu. cruzi, Lu. forattenii, Lu. longipalpis, Lu. migonei, Lu. olmeca, Lu. ovallesi, Lu. pseudolongipalpis, Lu. sallesi) in the area of Cuzco also argue against L. infantum as the Leishmania species infecting the mummies.

Because of the multiplicity of hypotheses regarding the Leishmania species involved, it is crucial to identify accurately the Leishmania found in the digestive tract of the pre-Columbian mummies:

- i. On the one hand, the infection could be caused by a Leishmania species, usually known to be responsible for cutaneous forms, associated or not with concomitant infection or disease. It would be interesting to gather more information regarding the presence of parasitic, viral or bacterial coinfection(s) that may cause immunosuppression. The microbiome analyses should give us some leads.
- ii. On the other hand, these findings could be indicative of the presence of an active indigenous transmission cycle of AVL during the pre-Columbian period. The only other known species of Leishmania, which causes AVL in continental South America, endemic to this geographic area, is L. colombiensis. Its active transmission cycle may have occurred either in Cuzco or in the neighboring countries, which were, at that time, part of the Inca Empire. Although the proven vector for L. colombiensis is currently unknown, this species has been isolated in the endemic area from Lu. hartmanni and Lu. shannoni (Kreutzer et al. 1991; Ramírez et al. 2016), and is able to develop in colony-reared Lu. trapidoi (Kreutzer et al. 1991). Interestingly, the presence of Lu. hartmanni has been reported in the area of Cuzco (Sanchez-Saldaña et al. 2004), which adds further evidences supporting this hypothesis.
- iii. Another possibility about the origin and the identity of the infection in these pre-Columbian mummies would involve another Leishmania species not yet identified. The Amazon basin contains a large diversity of Leishmania species. Only a small proportion of them have been described as formal species. The current restrictions on the capture and sacrifice of sylvatic animals for research purposes and the difficulties to conduct field works make that most of this diversity remains underexplored. It is possible that further indigenous parasites, not yet identified, capable of causing VL could be present. Therefore, the discovery, in the area of Cuzco or in the neighboring Amazonian basin, of an active transmission cycle involving a Leishmania species, which possesses the capacity to disseminate into internal organs, cannot be ruled out.
- iv. The last hypothesis implies that the etiological agent of the VL identified in pre-Columbian mummies by Santiago-Rodriguez *et al.* (2016) was introduced in South America together with the dogs brought by early human migrants (Van Ash *et al.* 2013) from East Asia by 14.6 Kya/13 Kya (date of the split between Northern and Southern branches) or by these migrants themselves (Raghavan *et al.* 2015). This agent could

be related to *L. donovani* or *L. infantum*. Nevertheless, traces of such event have not been found in genetic analyses of isolates from South American patients (Lukes *et al.* 2007).

Therefore, studies performed on pre-Columbian mummies require the identification of the exact causative agent (if not *L. infantum/L. chagasi*). It could be rather related to an indigenous species and/or to *Leishmania* parasites whose identity and origin have to be investigated. This would not be incompatible with the common hypothesis of an introduction of *L. infantum* through the Iberian conquest but would simply imply that *Leishmania* species with visceralization capabilities were already present before the arrival of the Spanish and Portuguese conquistadores.

QUEST FOR ANCIENT LEISHMANIA DNA IN HUMAN REMAINS

Ancient microbial DNA (amDNA) like ancient DNA (aDNA) fragments are characterized by highly fragmented DNA (more or less 100 nt) and DNA damages such as cytosine deaminated DNA molecules (Dabney, Meyer and Pääbo 2013). In the aDNA field, such short DNA fragments are often present in environmental DNA samples which make difficult the authentication of DNA fragments extracted from ancient remains. Before the emergence of the NGS, a very stringent procedure, including a replication of the results in an independent laboratory, was used to identify the genetic material. Clearly, the advent of NGS allowed for a massive output of aDNA sequences, giving the opportunity to explore in more detail the features of ancient DNA fragments like deamination toward the ends of the fragments (Briggs et al. 2007; Overballe-Petersen, Orlando and Willerslev 2012). But three things complicate the analysis of amDNA: the short length of aDNA fragments, the possible lack of reference genomes to match with and the fact that there is a highsequence similarity within and among taxa when short genetic sequences are used. The genus Leishmania is a slow evolving monophyletic group (Marcili et al. 2014). Comparative genomic analysis of Leishmania revealed divergence at the gene or pseudogene level with genes that are restricted to individual species (Peacock et al. 2007).

The colonization of ancient samples after the individual's death until its excavation or laboratory contamination could confound the analysis. Laboratory contamination by *Leishmania* DNA is unlikely to happen without the presence of *Leishmania* cell cultures and/or purified *Leishmania* DNA during samples processing. The vector of transmission of *Leishmania* to their hosts is through insect bite. Therefore, potential contamination of human remains, after the death, is not possible because *Leishmania* has no ability to survive out of its vertebrate (mammals, lizards, etc.) or invertebrate (Phlebotominae sand fly) hosts.

The use of amDNA to address the evolutionary history of the Leishmania genera and subgenera will benefit from NGS advances. To perform such analyses, it will be necessary to have more reference genomes, representative of the currently known diversity of the Leishmania genera and subgenera. For instance, 16 complete genomes representative of 14 Leishmania species are currently available to perform such analysis (http://tritrypdb. org/tritrypdb/). The majority of these genomes refer to species belonging to the Leishmania subgenus of the Euleishmania section. Within the Paraleishmania section, only Endotrypanum monterogeii (strain LV88) was sequenced. First, it is now essential to have more reference genomes to implement NGS metagenomic accurate assignation of ancient Leishmania DNA. Second, the use of appropriate molecular markers on which a target-enrichment strategy is amenable (Schuenemann et al. 2011) will help to perform confident Leishmania species assignation (Akhoundi et al. 2017).

Because of the diversity encompassed by leishmaniases (including a spectrum of diseases caused by more than 20 Leishmania species), the type of tissue sampled for the analysis will address different questions. CL, the most common Leishmania symdrome, affects the skin with lesions often occurring in the area of the bites of infected sand flies. But a minority of cutaneous infections caused by species of the Vianna subgenus is associated with concomitant or late ML and can result in destructive lesions of the nasopharyngeal/laryngeal mucosa (Aronson et al. 2016). These types of lesions are often observed in mummies of the pre-Columbian era (see Novo et al. 2016). Therefore, the body areas exposed to the bites of sand flies, i.e the hand, the face, the ears, the legs, and so on , are most likely to contain ancient Leishmania DNA. Unlike CL, VL and AVL do not produce visible lesions. VL shows the dissemination of Leishmania parasites throughout the reticulo-endothelial system and their ability to colonize and replicate into various internal organs (spleen, liver, bone marrow, etc.) (see for review Akhoundi et al. 2016; Aronson et al. 2016).

CONCLUSIONS

Modern techniques aimed at identifying ancient pathogen DNA not only help to identify a disease but also provide information on its frequency and evolutionary origin. They can also trace back sociocultural contacts and their potential role in the transmission and spread of infectious diseases. These techniques have successfully detected Leishmania infections in ancient Egyptian and Nubian mummies. They also gave crucial information on contacts between these two populations during the Middle Kingdom of Pharaonic Egypt (2050-1650 BC) (Zink et al. 2006) (See Fig. 2). This methodology showed the occurrence of visceral leishmaniasis in remains of Eleonora from Toledo, the wife of Cosimo I de Medicis, a famous person of the Italian Renaissance (Nerlich et al. 2012). They have also revealed some unconventional Leishmania infections, like the presence of a DNA signature of a non-human pathogenic Leishmania species, namely L. tarentolae, in the rib, skin and abdominal regions of a 300 -year-old human Brazilian mummy (Novo et al. 2015). Finally, it is a way to highlight the past occurrence of leishmaniases in currently leishmaniasis-free areas, but the identification of the exact species was not performed (Costa et al. 2009).

Based on what is known about the evolutionary history and epidemiology of AVL, the findings presented by Santiago-Rodriguez *et al.* (2016) emphasize the interest of studying pathogens causing infectious diseases in the light of ancient DNA. It opens new perspectives on the pre-Columbian *Leishmania* epidemiology, the clinical forms present during the pre-Columbian period and the identity of the causative agents.

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AUTHORS' CONTRIBUTIONS

DS and PP drafted the manuscript. All finalized the manuscript. All authors read and approved the final version of the manuscript.

Conflict of interest. None declared.

REFERENCES

- Akhoundi M, Downing T, Votypka J et al. Leishmania infection: molecular target and diagnosis. Mol Aspects Med 2017, In press.
- Akhoundi M, Kuhls K, Cannet A *et al*. A historical overview of the classification, evolution, and dispersion of *Leishmania* parasites and sandflies. PLoS Neglect Trop D 2016;**10**:e0004349.
- Alvar J, Aparicio P, Aseffa A et al. The relationship between leishmaniasis and AIDS: the second 10 years. Clin Microbiol Rev 2008;21:334–59.
- Alvar J, Vélez ID, Bern C et al. WHO leishmaniasis control team. Leishmaniasis worldwide and global estimates of its incidence. PLoS One 2012;7:e35671.
- Aronson N, Herwaldt BL, Libman M et al. Diagnosis and treatment of leishmaniasis: clinical practice guidelines by the Infectious Diseases Society of America (IDSA) and the American Society of Tropical Medicine and Hygiene (ASTMH). Clin Infect Dis 2016;63:e202–64.
- Barral A, Badaro R, Barral-Netto M et al. Isolation of Leishmania mexicana amazonensis from the bone marrow in a case of American visceral leishmaniasis. Am J Trop Med Hyg 1986;35:732–4.
- Barral A, Pedral-Sampaio D, Grimaldi, Jr et al. Leishmaniasis in Bahia, Brazil: evidence that Leishmania amazonensis produces a wide spectrum of clinical diseases. Am J Trop Med Hyg 1991;44:536–46.
- Barratt J, Kaufer A, Peters B et al. Isolation of novel Trypanosomatid, Zelonia australiensis sp. nov. (Kinetoplastida: Trypanosomatidae) provides support for a gondwanan origin of dixenous parasitism in the leishmaniinae. PLoS Neglect Trop D 2017;11:e0005215.
- Barro-Traoré F, Preney L, Traoré A et al. Cutaneous leishmaniasis due to Leishmania major involving the bone marrow in an AIDS patient in Burkina Faso. Ann Dermatol Venereol 2008;**130**:380–3.
- Briggs AW, Stenzel U, Johnson PL et al. Patterns of damage in genomic DNA sequences from a Neandertal. P Natl Acad Sci USA 2007;104:14616–21.
- Cnudde F, Raccurt C, Boulard F et al. Diffuse cutaneous leishmaniasis with visceral dissemination in an AIDS patient in Guadeloupe, West Indies. AIDS 1994;**8**:559–60.
- Costa MA, Matheson C, Iachetta L et al. Ancient leishmaniasis in a highland desert of Northern Chile. PLoS One 2009;4:e6983.
- Croan DG, Morrison DA, Ellis JT. Evolution of the genus Leishmania revealed by comparison of DNA and RNA polymerase gene sequences. Mol Biochem Parasitol 1997;89: 149–59.
- Cupolillo E, Medina-Acosta E, Noyes H et al. A revised classification for Leishmania and Endotrypanum. Parasitol Today 2000;16:142–4.
- Dabney J, Meyer M, Pääbo S. Ancient DNA damage. Cold Spring Harb Perspect Biol 2013;5:7.
- Delgado O, Castes M, White AC, Jr et al. Leishmania colombiensis in Venezuela. Am J Trop Med Hyg 1993;**48**:145–7.
- Espinosa OA, Serrano MG, Camargo EP et al. An appraisal of the taxonomy and nomenclature of trypanosomatids presently classified as *Leishmania* and *Endotrypanum*. *Para*sitology 2016;**15**:1–13.
- Fishman JA. Infections in immunocompromised hosts and organ transplant recipients: essentials. *Liver Transplant* 2011;17:S34–7.
- Floral R, Aghazadzh-Dibavar S, Bandyopadhyay M et al. Immunosuppression during Leishmania donovani infection: a

potential target for the development of therapy. Ann Parasitol 2014;60:239–45.

- Geiger A, Bossard G, Sereno D et al. Escaping deleterious immune response in their hosts: lessons from Trypanosomatids. Front Immunol 2016;7:212.
- Gerszten E, Allison M, Maguire B. Paleopathology in South American mummies: a review and new finding. *Pathobiology* 2012;**79**:247–56.
- Harkins KM, Schwartz RS, Cartwright RA et al. Phylogenomic reconstruction supports supercontinent origins for Leishmania. Infect Genet Evol 2016;**38**:101–9.
- Hernández D, Rodríguez N, Martínez C et al. Leishmania braziliensis causing visceral leishmaniasis in a patient with human immunodeficiency virus infection, identified with the aid of the polymerase chain reaction. Trans R Soc Trop Med Hyg 1993;**87**:627–8.
- Hernández DE, Rodriguez N, Wessolossky M et al. Visceral leishmaniasis due to a Leishmania variant that shares kinetoplast DNA sequences with Leishmania braziliensis and Leishmania mexicana in a patient infected with human immunodeficiency virus: identification of the Leishmania species with use of the polymerase chain reaction. Clin Infect Dis 1995;21: 701–2.
- Kerr SF, Merkelz R, MacKinnon C. Further support for a Palaearctic origin of Leishmania. Mem Inst Oswaldo Cruz 2000;95: 579–81.
- Kreutzer RD, Corredor A, Grimaldi G, Jr et al. Characterization of Leishmania colombiensis sp. (Kinetoplastida: Trypanosomatidae), a new parasite infecting humans, animals and phlebotomine sandflies in Colombia and Panama. Am J Trop Med Hyg 1991;446:662–75.
- Kuhls K, Alam MZ, Cupolillo, E et al. Comparative microsatellite typing of new world Leishmania infantum reveals low heterogeneity among populations and its recent old world origin. PloS Neglect Trop D 2011;5:e1155.
- Leblois R, Kuhls K, François O et al. Guns, germs and dogs: the origin of Leishmania chagasi. Infect Genet Evol 2011;11: 1091–5.
- Lukes J, Mauricio IL, Schönian G et al. Evolutionary and geographical history of the *Leishmania donovani* complex with a revision of current taxonomy. P Natl Acad Sci USA 2007;**104**: 9375–80.
- Marcili A, Sperança MA, da Costa AP et al. Phylogenetic relationships of Leishmania species based on trypanosomatid barcode (SSU rDNA) and gGAPDH genes: taxonomic revision of Leishmania (L.) infantum chagasi in South America. Infect Genet Evol 2014;**25**:44–51.
- Mauricio IL, Stothard JR, Miles MA. The strange case of Leishmania chagasi. Parasitol Today 2000;6:188–9.
- Momen H, Grimaldi G, Jr, Deane LM. Leishmania infantum, the etiological agent of American visceral leishmaniasis (AVL)? Mem Inst Oswaldo Cruz 1987;82:447–8.
- Nerlich AG, Bianucci R, Trisciuoglio A et al. Visceral leishmaniasis during Italian Renaissance, 1522-1562. Emerg Infect Dis 2012;**18**:184–6.
- Novo SP, Leles D, Bianucci R et al. Leishmania tarentolae molecular signatures in a 300 hundred-years-old human Brazilian mummy. Parasit Vector 2015;**4**:72.

- Novo SP, Leles D, Bianucci R et al. The process of Leishmania infection-disease and new perspectives of paleoparasitology. *Rev Inst Med Trop SP* 2016;**48**:45.
- Noyes HA, Morrison DA, Chance ML et al. Evidence for a neotropical origin of Leishmania. Mem Inst Oswaldo Cruz 2000;95:575–8.
- O'Dea A, Harilaos AL, Coates AG et al. Formation of the isthmus of Panama. Sci Adv 2016;**2**:e1600883.
- Overballe-Petersen S, Orlando L, Willerslev E. Next-generation sequencing offers new insights into DNA degradation. *Trends Biotechnol* 2012;**30**:364–8.
- Peacock CS, Seeger K, Harris D *et al*. Comparative genomic analysis of three Leishmania species that cause diverse human disease. Nat Genet 2007;**39**:839–47.
- Raghavan M, Steinrücken M, Harris K et al. Population genetics. Genomic evidence for the Pleistocene and recent population history of native Americans. *Science* 2015;**349**:aab3884.
- Ramírez JD, Hernández C, León CM et al. Taxonomy, diversity, temporal and geographical distribution of cutaneous leishmaniasis in Colombia: a retrospective study. Sci Rep 2016;6:28266.
- Ramos-Santos C, Hernandez-Montes O, Sanchez-Tejeda G et al. Visceral leishmaniosis caused by Leishmania (L.) mexicana in a Mexican patient with human immunodeficiency virus infection. Mem Inst Oswaldo Cruz 2000;95:733–7.
- Ready PD. Epidemiology of visceral leishmaniasis. Clin Epidemiol 2014;6:147–54.
- Rodriguez-Bonfante C, Bonfante-Garrido R, Grimaldi G, Jr et al. Genotypically distinct Leishmania colombiensis isolates from Venezuela cause both cutaneous and visceral leishmaniasis in humans. Infect Genet Evol 2003;3:119–24.
- Sanchez-Saldaña DL, Sáenz-Anduaga E, Pancorbo-Mendoza J et al. Leishmaniasis. Dermatología Peruana 2004;4:82–98.
- Santiago-Rodriguez TM, Fornaciari G, Luciani S et al. Taxonomic and predicted metabolic profiles of the human gut microbiome in pre-Columbian mummies. FEMS Microbiol Ecol 2016;**92**, DOI: 10.1093/femsec/fiw182.
- Sarkari B, Bavarsad Ahmadpour N, Moshfe A et al. Molecular evaluation of a case of visceral leishmaniasis due to *Leishmania tropica* in Southwestern Iran. Iran J Parasitol 2016;11: 126–30.
- Schuenemann VJ, Bos K, DeWitte S et al. Targeted enrichment of ancient pathogens yielding the pPCP1 plasmid of Yersinia pestis from victims of the Black Death. P Natl Acad Sci USA 2011;**108**:E746–52.
- Spotin A, Parvizi P. Comparative study of viscerotropic pathogenicity of *Leishmania major* amastigotes and promastigotes based on identification of mitochondrial and nucleus sequences. *Parasitol Res* 2016;**115**:1221–8.
- Van Ash B, Zhang AB, Oskarsson MC et al. Pre-Columbian origins of native American dog breeds, with only limited replacement by European dogs, confirmed by mtDNA analysis. Proc Biol Sci 2013;280:20131142.
- Zijlstra EE. PKDL and other dermal lesions in HIV co-infected patients with leishmaniasis: review of clinical presentation in relation to immune responses. PLoS Neglect Trop D 2014;8:e3258.
- Zink AR, Spigelman M, Schraut B et al. Leishmaniasis in ancient Egypt and Upper Nubia. Emerg Infect Dis 2006;**12**:1616–7.