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Natural infection by *Leishmania infantum* in the *Lutzomyia longipalpis* population of an endemic coastal area to visceral leishmaniasis in Brazil is not associated with bioclimatic factors

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# Abstract

Visceral leishmaniasis (VL) is a zoonosis caused by the protozoan *Leishmania infantum* and in Brazil is transmitted mainly by the bite of Lutzomuyia longipalpis sand flies. Data about the presence, distribution, natural infection rate, seasonal and monthly dynamics of the vector population are important for optimizing the measures to control VL in endemic areas. This study aimed to identify sand fly fauna in an endemic area for VL to detect the prevalence of L. infantum infection in the Lu. longipalpis population and to elucidate the influence of bioclimatic factors on the monthly fluctuations of this vector. HP light traps were monthly set in the intradomicile and peridomicile of residences located in the central and beachfront areas of Camacari, a VL endemic area. The sand fly collection was conducted in two periods: i) period 1-between December 2011 and November 2012 and ii) period 2-August 2014 and July 2015. Sand fly species were identified and detection of L. infantum infection by qPCR was performed in pools of female Lu. longipalpis. For the first time, the parasite load of positive pools was correlated with the number of Lu. longipalpis captured per month in both periods. Correlation analyses between the monthly fluctuation of the sand fly population and bioclimatic indices of the municipality in both collection periods were also performed. In both evaluated periods, more than 98% of the collected sand flies were Lu. longipalpis, confirming the predominance of this species in the region. It was captured mostly in the beachfront area in all months evaluated (99%). For the period 1, Leishmania DNA was detected in 81% of tested pools representing a minimal infection rate of 9.6%. In the period 2, 40% of the pools were positive with a minimal infection rate of 10.2%. Infected sand flies were only detected in the beachfront area in both periods. The parasite load was low and did not vary in the evaluated months despite the number of collected sand flies. No

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correlation was observed for climatic factors in both areas of Camaçari. These findings emphasize the high risk of *Leishmania* transmission in Camaçari regardless of the season and that other factors, aside from bioclimatic elements, are influencing the sand fly population monthly fluctuation.

#### Author summary

Visceral leishmaniasis (VL) is a parasitic disease transmited by a sand fly vector. VL is an important public health issue in Brazil occurring in all its regions with increasing incidences in big urban centers. In the State of Bahia, VL has spread to new hotspots in the last decade reaching the coastal area to the north of Bahia State capital, Salvador. This region underwent several environmental changes due to a rapid urbanization and construction of touristic and vacation complexes. Our investigation represents the first entomological study conducted in Camaçari. We evaluated the prevalence of the parasite infection in the sand fly population in two periods, between December 2011 and November 2012 and between August 2014 and July 2015. Our findings indicate that VL transmission occurs during the whole year and support the idea that the vector is not sensible to bioclimatic variations in Camaçari. Additionally, the maintenance of the parasite infection in the vector regardless of the season, indicates that the disease is well stablished in Camaçari, especially in the coastal area. These results emphasizes the urgency for a rigorous entomological and epidemiological survey aiming to guide control strategies to decrease VL cases.

## Introduction

Visceral leishmaniasis (VL) is an infectious disease considered to be neglected by the World Health Organization, mainly affecting poor regions in developing countries [1]. Previously in Brazil, likewise in other countries, the occurrence of VL was limited to rural areas and small urban localities [2,3]. However, the disease underwent an urbanization process with high incidence in big urban centers such as Belo Horizonte (Minas Gerais State), Campo Grande (Mato Grosso do Sul State), Palmas (Tocantins State) and Araçatuba (São Paulo State) [2, 4, 5, 6]. Nowadays, approximately 70% of VL cases occur in urban areas [3]. In the last decade, canine and human cases of VL were diagnosed in the southern States of Brazil, considered non-endemic until 2008 [7, 8, 9, 10, 11].

In the State of Bahia, several studies were previously conducted, aiming to identify the phlebotomine sand fly species present in municipalities endemic for VL, such as Jacobina, Jequié and Camaçari [12, 13, 14]. Nevertheless, these studies are old, lacking in the literature updated data on the sand fly species present in Bahia. Additionally, little is known about the fauna of vectors in urban and peri-urban areas of municipalities in Bahia where the transmission is considered intense, as occurs in Camaçari municipality [15, 16]. Thus, entomological surveys should be conducted in these areas to elucidate which sand flies species are present, the density of the vector population, its distribution, and seasonality. Studies to evaluate the natural infection rate of *Lu. longipalpis* with *L. infantum* and to elucidate the seasonal and monthly population dynamics are also necessary. Such studies should be performed employing entomological monitoring and evaluation of climatic factors' influence on the sand fly population. These evaluations are essential for the optimization of effective control measures for VL to be implemented in endemic areas [17, 18, 19, 20, 21, 22, 23].

In the evaluation of captured sand flies, the classical microscopic analysis and culture methods have been used to detect natural infection in sand flies, these techniques were performed in several studies [24, 25, 26, 27, 28]. However, the low sensitivity of these methods, the operational difficulty and delay to deal with a large number of samples and the low prevalence of *L. infantum* infection in the vectors in endemic areas limit the detection of parasites in sand flies [29, 30, 31]. In this context, a series of PCR assays to detect *Leishmania* DNA has been applied in studies to determine the natural infection rates in sand flies in the last years. Such assays aim to replace the aforementioned classical methods to increase the sensitivity and specificity in the detection of *Leishmania* parasites in the vectors [32, 33, 34, 35, 36, 37].

The aim of the present work was to identify the sand fly fauna present in Camaçari in two collection periods and to determine, for the first time, the prevalence of *Leishmania* infection in the sand fly population of this municipality during the whole year and, additionally, to elucidate the influence of bioclimatic factors on the population dynamics of these insects.

## Materials and methods

#### **Ethics statement**

All experimentation involving canine specimens was performed in compliance with Brazilian federal law for animal experimentation (Law 11794), in conformity with the guidelines for animal experimentation established by the Oswaldo Cruz Foundation (FIOCRUZ), and in accordance with the procedures described in the Brazilian Ministry of Health's manual for the VL surveillance and control. The present study received approval from the Institutional Review Board (CEUA protocol no. 015/2009, 017/2010 and 007/2013) of the Institute Gonçalo Moniz in Bahia, Brazil (IGM–FIOCRUZ/BA). Dog owners who agreed to participate in this study signed a form of Free, Prior and Informed Consent (FPIC).

#### Study area and selection of households

The study was conducted in the municipality of Camaçari, considered the biggest in the metropolitan region of Salvador, located at latitude 12°52'30" and longitude 38°28'52". The climate in Camaçari is hot and humid, with minimum temperatures above 18°C, terrain formed by fluvial-marine and coastal plains and pre-coastal vegetation. Camaçari is an endemic area for visceral leishmaniasis and during the study two human cases occurred in Monte Gordo, a city located in the neighborhood of VL area evaluated [38]. For the entomological study, Camaçari was stratified into two areas: beachfront (close to the coastal strip) and central (far from the coastal strip) (Fig 1).

The collections of the insects were conducted in two one-year periods: i) period 1 from December 2011 to November 2012 and ii) period 2 –from August 2014 to July 2015. Captures were performed in neighborhoods presenting a high prevalence of canine visceral leishmaniasis (CVL). In period 1, the sand fly captures were done in 12 households (collection sites) distributed in three neighborhoods in the beachfront area: Monte Gordo (3 houses), Barra de Jacuípe (2 houses) and Jauá (1 house), and six neighborhoods in the central area (1 house in each one): Parque Real Serra Verde, Verde Horizonte, Nova Vitória, Parque Verde II, Phooc III, and Jardim Limoeiro. In period 2, the captures were performed in four neighborhoods: Machadinho in the central area, and Jauá, Barra de Jacuípe and Monte Gordo in the beachfront area. In each one, four households were selected, making a total of 16 collection sites.

In both collection periods, the residences that presented higher chances of the collection were selected, based on the presence of natural breeding sites for sand flies. The presence of

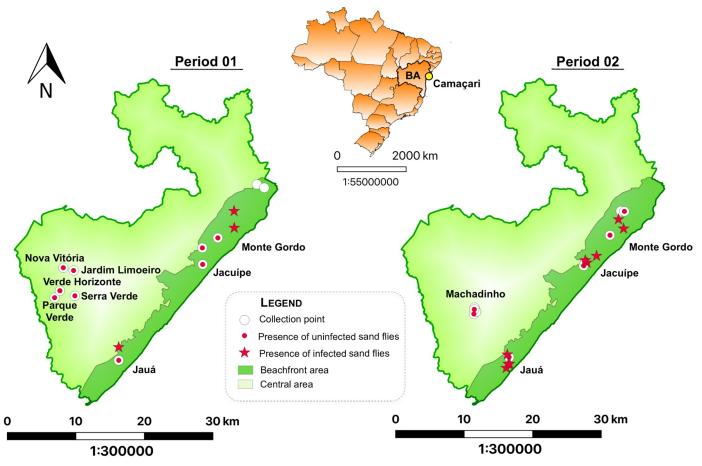


Fig 1. Map illustrating the localization of the Municipality of Camaçari in the state of Bahia, Brazil, and the sand fly collection points in the two one-year **periods**. Maps show the spatial distribution of sand fly collection points (white dots) in each period: the red dot indicating the presence of sand flies and the red star indicating the presence of infected sand flies. The beachfront area is represented as dark green (close to the coastal strip). Source: QGIS 2.14.3-Essen.

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abundant organic matter, areas with high humidity, animal shelters close to residences, visible tree roots and holes in stones or soil in the peridomicile were considered attractive factors for the presence of sand flies. Information regarding the occurrence of VL and/or CVL cases in the residence and neighboring houses, notified by the municipality's epidemiological survey in the last five years or informed by the residents were also taken into account.

## Collection, identification and design of sand fly pools

The investigations were carried out with light traps, which were placed every month until 12 months, at the intradomicile and peridomicile of households for three consecutive nights. The traps remained on between dusk (17:00h) and dawn (6:00h) of the next day when the insects were captured and gathered. The captured sand flies were identified as described by Young and Duncan [39], in which the males were clarified and mounted on slides while for the females, only the head and the last two abdominal segments were mounted in Berlese liquid, according to a modified technique from Langeron [40]. The rest of the female bodies were stored in 6% dimethyl sulfoxide (DMSO) and kept at—70°C until the design of pools and DNA extraction.

Each pool was composed by two to ten female fragments of the species *Lu. longipalpis* collected in the same neighborhood, month and household. A total of 674 *Lu. longipalpis* non-fed females were used to build 74 pools in period 1, and 166 females were divided into 42 pools in period 2.

#### Genomic material extraction

The pools of *Lu. longipalpis* females were macerated and homogenized in  $60\mu$ L of lysis buffer (100mM Tris-HCL, 100 mM NaCl, 25 mM EDTA, 0,5% SDS, pH 8,0) and  $10\mu$ L of proteinase k at 1 mg/L (Invitrogen Life Technologies, Carlsbad, CA, USA). The macerated material was incubated in a water bath for 16 to 18 hours at 37°C. The DNA was purified using phenol-chloroform technique and then precipitated with ethanol following the protocol described by Michalsky et al. (2002). The DNA samples were resuspended in 50 $\mu$ L TE buffer (Tris-EDTA 1X), and their concentrations were measured by digital spectrophotometry (Nanodrop ND-1000 Thermo Scientific, Wilmington, USA). Lastly, the samples were aliquoted at 20 ng/ $\mu$ L and stored at -20°C until their usage for qPCR.

#### qPCR assay

The amplification of the gene control of *Lu. longipalpis* (GenBank AF446142; [41]) was used to normalize the DNA concentration of *L. infantum* present in the sand fly pool samples, as well as to assure that the negative results did not occur due to DNA degradation or the presence of inhibitors. For the amplification of the gene control, forward (Fleb-F) and reverse (Fleb-R) primers, 5'-AATTTCTTTTCCTTAGGACCATCGA-3' and 5'-TAGGACATCTTCGGAA AATTGTTG-3' respectively, were used together with a fluorogenic probe 5'-AMTCCTCAS AGTCTTTGACTCCACGTTGGTT-3' aiming for the *Lu. longipalpis* periodicity gene which does not amplify *Leishmania* genomic DNA. The temperature conditions for the reaction were 50°C for 2min, 95°C for 10min and 40 cycles of 95°C for 15s and 60°C for 1min. A standard curve for the gene control was used for each reaction using DNA from pools of colony-reared males. For each sample, the quantity of gene control amplified DNA was determined by the comparison between the samples' and curves' Ct values. Only the samples with amplification for periodicity gene were used, to determine the *Leishmania* infection rate.

qPCR assays aiming to determine the quantity of the parasite's DNA in the pools of captured female sand flies were based on a protocol described by Francino et al. [42] adapted by Solcà et al. [43]. The values obtained in each reaction were normalized from a standard threshold to minimize variations. The same methodology described by Borja et al. [44] was used to determine the cut-off. The parasite load was expressed by the mean number of parasites per pool triplicate obtained in qPCR reaction. The mean of the parasite load detected in positive pools was calculated as the sum of parasite loads divided by the number of positive pools.

#### Calculation of natural infection rate

The *Leishmania* natural infection rate in the sand fly pools was expressed as a minimum infection rate (MIR), calculated as the ratio of the number of positive pools by the total specimens tested x 100, according to Paiva et al. [45].

#### Georeferencing and attainment of climatic data

A Global Positioning System (GPS) device, Garmin eTrex HC, was used to georefence the residences included in the study. The software QGIS was used to make the maps of collection sites. A thermo-hygro-anemometer was employed to attain microclimatic data of temperature and air humidity at the trap locations. These data were measured once per month at the household's peridomicile during the placement of the traps during the study.

# Influence of climatic variables on monthly fluctuation of *Lu. longipalpis* population

The influence of climatic variables on the monthly density fluctuation of *Lu. longipalpis* was evaluated along each one of both one-year periods on the beachfront and central areas. Neighborhoods that were highlighted by its higher sand fly density such as Monte Gordo, Barra de Jacuípe, Jauá and Parque Verde II in the first period of the collection were also evaluated isolatedly. Monthly mean values of temperature (°C), air humidity (%) and rainfall (mm3) referring to the first collection year were taken into account for the analysis.

## Statistical analysis

Data banks were created on Epi Info, comprising epidemiological, environmental, bioclimatic and sand fly density data. The correlation between the mean parasite load and the number of sand flies captured per month in each neighborhood was evaluated using Spearman coefficient, it was considered significant when p < 0.05. The correlation between the bioclimatic variables and sand fly population density was evaluated using the Spearman coefficient, it was considered significant when p < 0.05.

## Results

## Sand fly collection

A total of 7,116 sand flies were collected, being 5,745 in period 1 and 1,151 in period 2. Morphological analysis of the sand fly fauna allowed the species identification of 6,321 specimens. In period 1, five species were identified with the predominance of *Lu. longipalpis* (98%), followed by *Lutzomyia sallesi* (0.8%), *Lutzomyia evandroi* (0.4%), *Lutzomyia whitmani* (0.2%) and *Lutzomyia choti* (0.1%) and specimens belonging to cortelezzii complex (0.1%), only identified by genus. For the period 2, 1,002 sand flies were identified demonstrating the same predominance of *Lu. longipalpis* (99.9%) (Table 1).

The most significant proportion of the captured sand flies were registered at the beachfront area (Fig 2). In the period 1, 99.6% were captured at this area and 0.4% at the neighborhoods of the central area, while in the period 2, around 100% of the sand flies were captured at the beachfront area. Monte Gordo, Jauá and Barra de Jacuípe had the most significant proportion of collected specimens in period 1, 76%, 12%, and 9% respectively. For the period 2, the collections were also higher in Jauá (56%) and Barra de Jacuípe (39%).

Regarding the monthly fluctuation of *Lu. longipalpis* population in the studied area, the highest collection was observed in September 2012 (1109 sand flies) and the lowest in November 2012 (154 sand flies) in the period 1 (Fig 2A), whereas in the period 2 the highest density was observed in February 2015 (371 sand flies) and the lowest in September 2014 (111 sad flies) (Fig 2B). In respect to the sand flies endophilic and exophilic behavior, the sand flies were predominantly collected in the periodmicile in both collection periods (S1 Table).

## Natural infection of Lu. longipalpis

All pools tested positive for *Lu. longipalpis* periodicity control gene by qPCR. The presence of *L. infantum* DNA was detected in 60 pools among 74 evaluated, in the period 1, indicating natural infection in the sand fly population with a MIR of 9.6% at the beachfront area (Table 2).

Captured species	First period										Second period				
	Beachfront area			Central area							Beachfront area			Central area	
	MG	JC	JA	PR	VH	NV	PV	PH	JL	Total N (%)	MG	JA	JC	MA	Total N (%)
Lutzomyia longipalpis	3,999	482	604	22	2	2	108	1	15	5,235 (98.4)	43	564	394	-	1,001 (99.9)
Lutzomyia sallesi	-	-	11	-	2	31	-	-	-	44 (0.8)	-	-	-	-	-
Lutzomyia evandroi	16	-	1	-	3	-	1	-	-	21 (0.4)	-	-	-	-	-
Lutzomyia whitmani	-	-	1	-	-	-	8	-	1	10 (0.2)	-	-	-	-	-
Lutzomyia choti	2	-	-	-	-	2	1	-	-	5 (0.1)	-	-	-	1	1 (0.1)
Cortellezzi sp.	4	-	-	-	-	-	-	-	-	4 (0.1)	-	-	-	-	-
Total	4,021	482	617	22	7	35	118	1	16	5,319 (100)	43	564	394	1	1,002 (100)

#### Table 1. Number of identified sand flies stratified by neighborhoods of Camaçari-BA municipality in the first and second collection periods.

MG: Monte Gordo, JC: Barra de Jacuípe, JA: Jauá, PR: Parque Real Serra Verde, VH: Verde Horizonte, NV: Nova Vitória, PV: Parque Verde II, PH: Phooc III, JL: Jardim Limoeiro, MA: Machadinho.

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The neighborhoods that presented positivity for *L. infantum* in captured sand flies were Monte Gordo, Barra de Jacuípe and Jauá, located at the beachfront area (Fig 1). Monte Gordo presented the majority of sand flies captured and positive pools, but MIR in Barra do Jacuípe (14.3%) was higher than in Monte Gordo (9.3%).

For the period 2, 17 out of 42 pools tested positive for the presence of *L. infantum* DNA, distributed in Jauá, Barra do Jacuípe and Monte Gordo (Table 2), all located at beachfront area showing a MIR of 10.2%. Barra do Jacuípe presented the majority of positive pools, wherein the MIR was 13.3% lower than Monte Gordo (20.0)%. Natural infection was not detected in sand flies from the central area in both periods.

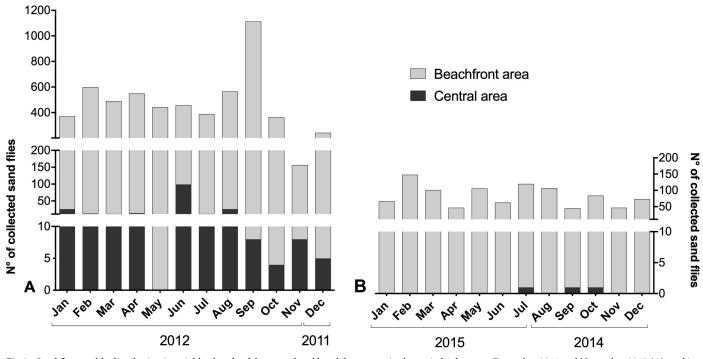


Fig 2. Sand fly monthly distribution in neighborhoods of the central and beachfront areas in the period 1, between December 2011 and November 2012 (A), and in the period 2, between August 2014 and July 2015 (B).

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Study	period		Monte Gordo			Barra de Jacuípe	Jauá			
		Sand flies	Pools +/Total	MIR	Sand flies	Pools +/Total	MIR	Sand flies	Pools +/Total	MIR
2011	Dec	23	3/3	13.0	-	-	-	-	-	-
2012	Jan	20	2/4	10.0	-	-	-	-	-	-
	Feb	38	5/5	11.4	7	1/1	14.3	-	-	-
	Mar	30	2/3	2/3 6.7		1/1	33.3	4	1/1	25.
	Apr	50	5/5 10.0		9	1/1	11.1	-	-	-
	May	43	3/5	7.0	-	-	-	4	0/1	-
	Jun	37	1/4	2.7	-	-	-	-	-	-
	Jul	3	1/1	33.3	-	-	-	-	-	-
	Aug	-	-	-	-	-	-	-	-	-
	Sep	218	20/23 9.2		-	-	-	10	0/1	-
	Oct	90	9/9	10.0	-	-		4	1/1	16.
	Nov	7	1/1	14.3	13	3/3	23.1	-	-	-
Total period 1		559	52/63	9.3	42	6/7	14.3	22	2/4	9.0
2014	Aug	-	-	-	10	1/2	10.0	14	0/1	7.1
	Sep	-	-	-	2	1/2	50.0	2	1/2	50
	Oct	-	-	-	3	1/1	33.3	6	1/4	16.
	Nov	-	-	-	28	3/7	10.7	2	0/1	-
	Dec	-	-	-	1	0/1	100.0	36	2/7	5.6
2015	Jan	1	0/1	-	9	0/2	-	3	0/1	-
	Feb	-	-	-	2	0/1	-	5	0/1	-
	Mar	-	-	-	-	-	-	10	0/1	-
	Apr	-	-	-	-	-	-	-	-	-
	May	-	-	-	1	1/1	100.0	3	1/1	33.
	Jun	9	2/2	22.2	4	1/1	25.0	2	1/1	50.
	Jul	-	-	-	-	-	-	13	1/1	7.7
Total p	period 2	10	2/3	20.0	60	8/18	13.3	96	7/21	7.3

Table 2. Lu. longipalpis positive pools and minimal infection rates distributed by month and neighborhoods of the beachfront area of Camaçari-BA municipality in both collection periods.

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The natural infection of *Lu. longipalpis* was evaluated in all months of the study, except for August 2012 in period 1. *Leishmania* infection was observed in sand fly pools in all evaluated months in period 1. In period 2, the positivity was detected in sand fly pools in May, June, July, August, September, October, November and December (Table 2).

The mean parasite load of the positive pools was constantly low during both studied periods as seen in Fig 3, only three out of 60 positive pools had higher parasite load compared to the remaining ones in Monte Gordo in the first period (February, May and December) and one pool out of 17 in Barra de Jacuipe in the second period (October). The number of sand flies captured per month did not correlate with the mean parasite load.

## Influence of bioclimatic factors on Lu. longipalpis population

No correlation was found between any climatic factors and *Lu. longipalpis* monthly density in both areas, neither in the aggregated analysis nor looking at specific sites (Fig 4).

#### Household characterization

There were 33% of households presenting previous occurrence of CVL cases in period 1 and 44% in period 2. In addition, 47% and 13% presented CVL cases in the neighboring houses in

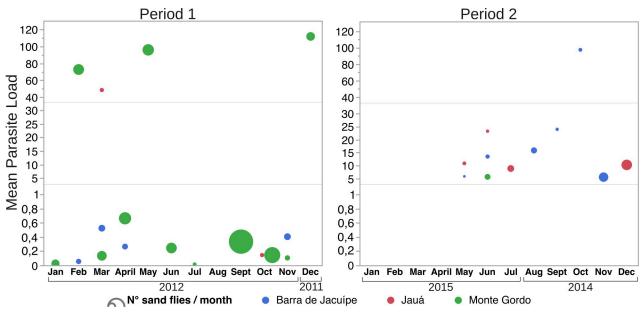


Fig 3. Mean parasite load detected in positive pools per month from each neighborhood. (A) in period 1; (B) in period 2. Dot size represents the number of sand flies captured.

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periods 1 and 2 respectively. In 60% of houses in period 1 someone in the past presented VL and 6.7% presented VL cases in the neighboring houses. In period 2, 6.3% of households presented previous VL cases in the same house and 18.8% in the neighboring houses.

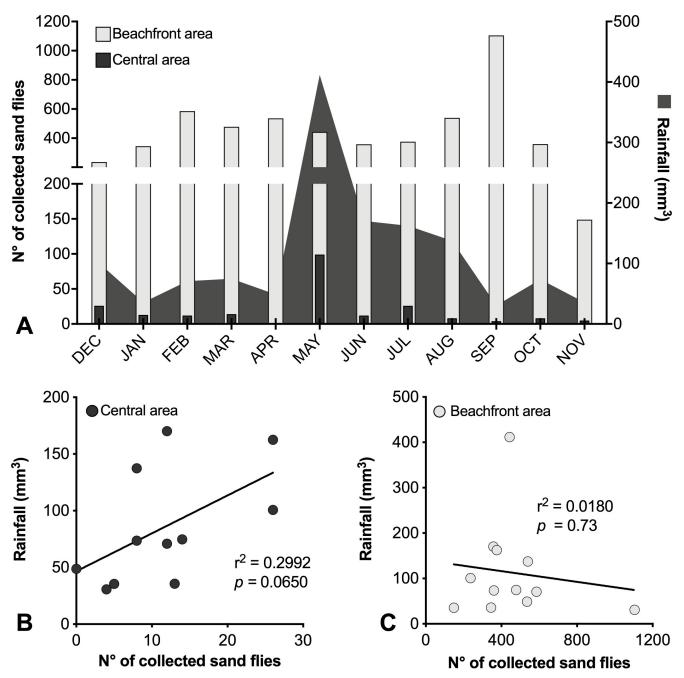
Households evaluated in both periods in central and beachfront area did not present statistical differences in their epidemiological characteristics. All of them presented vegetation in their backyard and surrounding area as well as organic matter, such as garbage, decomposing leaves and fruits, construction debris, animal feces and others. Thirty percent of houses in both periods did not have sewage system.

#### Discussion

The present study reported, in two moments, the predominance of the species *Lu. longipalpis* in the sand fly fauna of Camaçari, in which its natural infection with *L. infantum* was determined in neighborhoods of the beachfront area, and aimed to elucidate the influence of bioclimatic factors on the sand fly population.

Our findings demonstrate that although a diverse sand fly fauna, in period 1, has been registered in the investigated areas, the species *Lu. longipalpis* predominated in both investigated periods during all months of the study. These data are consistent with findings of similar entomological studies performed in other Brazilian endemic cities for visceral leishmaniasis [46, 47, 48]. *Lu. longipalpis* distribution in Camaçari demonstrated the predominance of this vector in neighborhoods located at the beachfront area, especially in the peridomicile of households in Monte Gordo and Jauá. The collection of *Lu. longipalpis* in areas close to the sea is an uncommon finding that has been poorly reported. Thus, it strengthens the idea that this vector has high adaptability to different biomes, being also present in environments such as coastal plains with pre-coastal vegetation which provides little subsidy for the formation of ecotopes for reproduction due its microclimatic characteristics [3, 16, 49, 50].

During period 1, two VL cases were identified in Monte Gordo (beachfront area), one case in 2011 and another one in 2012. There were vector control actions in Monte Gordo after VL



**Fig 4. Evaluation of the influence of monthly rainfall on sand flies monthly captured in the Central Area.** (A) Distribution of the mean monthly rainfall (mm3) concerning sand flies captured; (B) Correlation between the sand flies captured in central area and the mean monthly rainfall (mm3) (C) Correlation between the sand flies captured in beachfront area and the mean monthly rainfall (mm3) ( $r^2$ , p: Spearman coefficient).

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cases, resulting in a reduction of vector population. The difference observed in Table 1, when the density of captured *Lu. longipalpis* in Monte Gordo was lower in the second period in comparison with the first one, should have occurred due the vector control. Additionally, there was a chikungunya outbreak in Camaçari during period 2, which lead to the usage of insecticide spraying to reduce mosquitos that might have influenced the lower collection of sand flies in this period when compared to period 1.

In the present study, it is worth highlighting the capture of *Lu. sallesi* due to its importance related to the capability to be naturally infected with *L. infantum*, as shown in other studies, suggesting the need for monitoring this species in Camaçari [35]. Other authors emphasize the importance of constant monitoring of naturally infected sand fly species, whether they are known vectors or novel species involved in the transmission of leishmaniasis in endemic areas [28, 33, 35, 51, 52, 53, 54].

Our results demonstrated for the first time the prevalence of *L. infantum* infection in the *Lu. longipalpis* population in Camaçari. The MIR found in the first and second period (9.6 and 10.2%, respectively) corroborates to the findings of other studies, which demonstrated rates ranging from 0.2% to 10.7% in sand fly populations from other visceral leishmaniasis endemic areas [55, 56, 57, 58, 59, 60, 61]. MIR was similar in both periods demonstrating the sustainability of *Lu. longipalpis* populations potentially capable of infecting susceptible reservoirs, over the years in Camaçari municipality. MIR were higher than other studies [56, 62, 63] may be due to the higher sensitivity of the qPCR in comparison with the convenctional PCR used in previous entomological studies [30, 60, 64]. Such higher MIR was also detected using *Leishmania* kDNA-qPCR in a recent study, which detected a MIR of 65% in pools of engorged *Lu. longipalpis* females colleted in Rio Verde de Mato Grosso municipality [65]. Thus, a qPCR-based approach, used in the present work, may lead to a MIR closer to the *L. infantum* true prevalence in the *Lu. longipalpis* population, due to its higher sensitivity.

The identification of female *Lu. longipalpis* in all seasons during both collection periods, points out to the possible occurrence of transmission during the whole year, corroborating studies conducted in other endemic areas under distinct conditions [66, 67]. Moreover, throughout the study, in both periods, the parasite load was low and did not vary in the evaluated months despite the number of collected sand flies. Thus, this finding suggests that the risk of infection is the same during the whole year regardless of sand fly density.

Detection of *L. infantum* infection in samples from the *Lu. longipalpis* population, high prevalences of CVL and VL cases in Camaçari was observed in the same areas evaluated in two different moments. In both periods of the study, the highest CVL prevalences, the occurrence of VL cases, the number of collected sand flies and infection rate of vectors was observed in locations at the beachfront area. These findings corroborated with other prevalence studies [15, 16] and showed that the coastal area of Camaçari presents a high risk of CVL and VL.

The most significant proportion of captured *Lu. longipalpis* with evidence of natural infection was detected in the beachfront area, especially in Monte Gordo, Barra de Jacuípe and Jauá in contrast with the central area. This finding was not related to climatic factors or household characteristics such as presence of vegetation, hygiene conditions in the peridomicile and breeding of livestock and/or domestic animals. In this study the factors related to this difference was not identified. A more significant proportion of *Lu. longipalpis* was detected in the peridomicile, which is in accordance with results observed by other authors [17, 18, 20, 66, 68, 69]. Households studied in the first period were not the same as in the second period, as well as localities representing central area, but this difference possibly did not interfere in the analysis comparing central and beachfront areas. This is due to absence of statistical differences in household characterization between them comparing both periods.

The influence of climatic factors on *Lu. longipalpis* population was evaluated in the present study during the first collection period, and our results indicated that in the study area sand fly population density was not influenced by temperature, humidity and rainfall variations. Several authors found a correlation with climatic factors, observing a higher prevalence of *Lu. longipalpis* during the rainy season [70, 71, 72, 73, 74], and other ones demonstrated a preference of this species to dry and rocky areas [75, 76, 77]. The lack of correlation among climatic factors and sand fly density in the beachfront area, where more than 96% of sand flies were

captured, and the limited correlation found in central area indicated that other factors could be more important in modulating the sand fly population than climatic factors in the area studied. This finding probably occurs because in this area temperature and humidity does not vary along the year.

Regarding the temperature, there was no significant effect of its variation on *Lu. longipalpis* population, not being this fact a determinant factor of this species in monthly fluctuation in this municipality, as demonstrated by other authors concerning other regions of Brazil [66, 70, 78]. It is worth to highlight that the annual temperature cycle in Camaçari has a small range of variations, in average between 23°C and 27°C, which can justify the absence of correlation of this climatic factor. In other studies, similar results were found in other northeastern municipalities where the annual temperature cycle has likewise a small range of variation [79, 80]. In localities with such small temperature variation, changes in the rainfall and humidity should have a larger influence on the fluctuation of *Lu. longipalpis* population, as demonstrated by other authors' findings [72, 73, 74, 80]. However, our findings did not found humidity influence, possibly because humidity also has a small range of variation in Camaçari as well as coastal area, humidity remains high during the whole year.

Additionally, our more significant proportion of *L. infantum* positive sand flies found in the beachfront area denies what was proposed by Nieto and collaborators [76] niche modeling. It was shown in their work that the coastal area of Bahia has a lower risk for visceral leishmaniasis when compared to the central Caatinga region. Curiously, there was an outbreak of the disease in Camaçari after this study with both canine and human cases [16, 22]. The expansion to coastal areas has occurred due to recent environmental changes that were done in the beachfront of Camaçari for the construction of touristic and vacation complexes. The rapid urbanization of this location brought new human inhabitants who are used to breed dogs, chickens, and livestock in the peridomicile, providing breeding sites and blood source for *Lu. longipalpis* population.

In sum, there was no correlation between bioclimatic factors and sand fly density, being *Lu. longipalpis* the most prevalent sand fly species in Camaçari during both studied periods. Furthermore, the parasite load in *Lu. longipalpis* was predominantly low and did not fluctuate during the whole year despite the number of collected sand flies. These findings emphasize the high risk of *Leishmania* transmission in Camaçari regardless of the season and point out to the urgency for vector control actions, diminishing the number of CVL and VL cases especially in the neighborhoods in the municipalities beachfront area.

## **Supporting information**

S1 Table. The density of sand flies captured in each household's investigated sector according to the identified species in Camaçari-BA municipality between December 2011 and November 2012 and between August 2014 and July 2015. (DOCX)

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#### References

- 1. WORLD HEALTH ORGANIZATION. WHO. 2010. Research to support the elimination of visceral leishmaniasis. Available in: <a href="https://apps.who.int/iris/bitstream/handle/10665/70700/TDR\_BL10.10\_eng">https://apps.who.int/iris/bitstream/handle/10665/70700/TDR\_BL10.10\_eng</a>. pdf;jsessionid=377EE86E752F5D1549F9D13BBA109CA6?sequence=1>
- Werneck GL, Costa CHN, de Carvalho FAA, Pires e Cruz M do S, Maguire JH, Castro MC. Effectiveness of Insecticide Spraying and Culling of Dogs on the Incidence of Leishmania infantum Infection in Humans: A Cluster Randomized Trial in Teresina, Brazil. Warburg A, editor. PLoS Negl Trop Dis. 2014 Oct 30; 8(10):e3172. https://doi.org/10.1371/journal.pntd.0003172 PMID: 25357122
- Dos Reis LL, Balieiro AA da S, Fonseca FR, Gonçalves MJF. Changes in the epidemiology of visceral leishmaniasis in Brazil from 2001 to 2014. Rev Soc Bras Med Trop. 2017 Sep; 50(5):638–45. https:// doi.org/10.1590/0037-8682-0243-2017 PMID: 29160510
- Carranza-tamayo CO, Carvalho M do SL, Bredt A, Bofil MIR, Rodrigues RMB, Silva AD da, et al. Autochthonous visceral leishmaniasis in Brasília, Federal District, Brazil. Rev Soc Bras Med Trop. 2010; 43(4):396–9. PMID: 20802938
- Silva DA, Madeira MF, Teixeira AC, de Souza CM, Figueiredo FB. Laboratory tests performed on Leishmania seroreactive dogs euthanized by the leishmaniasis control program. Vet Parasitol. 2011 Jun; 179 (1–3):257–61. https://doi.org/10.1016/j.vetpar.2011.01.048 PMID: 21349644
- Soares MRA, Mendonça IL de, Bonfim JM do, Rodrigues JA, Werneck GL, Costa CHN. Canine visceral leishmaniasis in Teresina, Brazil: Relationship between clinical features and infectivity for sand flies. Acta Trop. 2011 Jan; 117(1):6–9. https://doi.org/10.1016/j.actatropica.2010.08.015 PMID: 20816657
- Steindel M, Menin Á, Evangelista T, Stoco PH, Marlow MA, Fleith RC, et al. Outbreak of autochthonous canine visceral leishmaniasis in Santa Catarina, Brazil. Pesqui Veterinária Bras. 2013 Apr; 33(4):490– 6.
- 8. Bianchi MV, Fredo G, Tagliari NJ, Filho RVL, De Lorenzo C, Lupion CG, et al. Autochthonous canine visceral leishmaniasis in urban area of Porto Alegre, RS, Brazil | Leishmaniose visceral canina autóctone na região urbana de Porto Alegre, RS, Brasil. Acta Sci Vet. 2016; 44:1–4.
- Teixeira MC, Stobbe NS, Schmidt V, Lima VMF de, Tartarotti AL, Ramos RR, et al. Seroepidemiology of Leishmania infantum in dogs in the city of Porto Alegre, Rio Grande do Sul, Brazil. Semin Ciências Agrárias. 2016 Dec 14; 37(6):4077.
- Trench FJP, Ritt AG, Gewehr TA, De Souza Leandro A, Chiyo L, Rittgewehr M, et al. First Report of Autochthonous Visceral Leishmaniosis in Humans in foz Do Iguaçu, Paraná State, Southern Brazil. Ann Clin Cytol Pathol. 2016; 2(6):1–4.

- Riboldi E, Carvalho F, Romão PRT, Barcellos RB, Bello GL, Ramos RR, et al. Molecular Method Confirms Canine Leishmania Infection Detected by Serological Methods in Non-Endemic Area of Brazil. Korean J Parasitol. 2018 Feb 28; 56(1):11–9. <u>https://doi.org/10.3347/kjp.2018.56.1.11</u> PMID: 29529845
- DEANE, L. M. Leishmaniose Visceral no Brasil. Estudos sobre reservatórios e transmissores realizados no Estado do Ceará.—Faculdade de Medicina, Universidade de São Paulo, São Paulo. Tese (Livre Docência), p. p. 143, 1956.
- Sherlock IA 1964. Surto de Calazar na Zona Central do Estado da Bahia. Rev Brasil Malar Doenças Trop 16: 157–170.
- 14. Sherlock IA 1969. Observações sobre calazar em Jacobina, Bahia. I—Histórico e dados preliminares. Rev Brasil Malar Doenças Trop 21: 523–534.
- Cunha S, Freire M, Eulalio C, Critosvao J, Netto E, Johnson WD, et al. Visceral leishmaniasis in a new ecological niche near a major metropolitan area of Brazil. Trans R Soc Trop Med Hyg. 1995 Mar; 89 (2):155–8. https://doi.org/10.1016/0035-9203(95)90474-3 PMID: 7778139
- Julião FS, Souza BMPS, Freitas DS, Oliveira LS, Larangeira DF, Dias-Lima AG, et al. Investigação de áreas de risco como metodologia complementar ao controle da leishmaniose visceral canina. Pesqui Vet Bras. 2007; 27(8):319–24.
- 17. Barata RA, França-Silva JC, Mayrink W, Silva J, Prata A, Lorosa ES, et al. Aspectos da ecologia e do comportamento de flebotomíneos em área endêmica de leishmaniose visceral, Minas Gerais Aspects of the ecology and behaviour of phlebotomines in endemic area for visceral leishmaniasis in State of Minas Gerais. Rev Soc Bras Med Trop. 2005; 38(5):421–5. PMID: 16172760
- Oliveira AG, Galati EAB, Fernandes CE, Dorval MEC, Brazil RP. Seasonal variation of Lutzomyia longipalpis (Lutz & Neiva, 1912) (Diptera: Psychodidae: Phlebotominae) in endemic area of visceral leishmaniasis, Campo Grande, state of Mato Grosso do Sul, Brazil. Acta Trop. 2008 Jan; 105(1):55–61. https://doi.org/10.1016/j.actatropica.2007.09.008 PMID: 18022137
- Michalsky ÉM, Fortes-Dias CL, França-Silva JC, Rocha MF, Barata RA, Dias ES. Association of Lutzomyia longipalpis (Diptera: Psychodidae) population density with climate variables in Montes Claros, an area of American visceral leishmaniasis transmission in the state of Minas Gerais, Brazil. Mem Inst Oswaldo Cruz. 2009 Dec; 104(8):1191–3. https://doi.org/10.1590/s0074-02762009000800020 PMID: 20140384
- Machado TO, Bragança MAL, Carvalho ML, Andrade Filho JD. Species diversity of sand flies (Diptera: Psychodidae) during different seasons and in different environments in the district of Taquaruçú, state of Tocantins, Brazil. Mem Inst Oswaldo Cruz. 2012 Nov; 107(7):955–9. <u>https://doi.org/10.1590/s0074-02762012000700021</u> PMID: 23147157
- Queiroz MFM, Varjão JR, Moraes SC de, Salcedo GE. Analysis of sand flies (Diptera: Psychodidae) in Barra do Garças, State of Mato Grosso, Brazil, and the influence of environmental variables on the vector density of Lutzomyia longipalpis (Lutz & Neiva, 1912). Rev Soc Bras Med Trop. 2012 Jun; 45 (3):313–7. PMID: 22760128
- 22. BRASIL. Ministério da Saúde. Manual de vigilância e controle da leishmaniose visceral. 1. ed. Brasília, DF, 2014, p. 120.
- Salomón OD, Feliciangeli MD, Quintana MG, Afonso MMDS, Rangel EF. Lutzomyia longipalpis urbanisation and control. Mem Inst Oswaldo Cruz. 2015 Oct 23; 110(7):831–46. https://doi.org/10.1590/0074-02760150207 PMID: 26517497
- Rangel EF, Souza NA de, Wermelinger ED, Barbosa AF. Infecção natural de Lutzomyia intermedia Lutz & Neiva, 1912, em área endêmica de leishmaniose tegumentar no Estado do Rio de Janeiro. Mem Inst Oswaldo Cruz. 1984; 79(3):395–6. https://doi.org/10.1590/s0074-02761984000300020 PMID: 6535924
- Feliciangeli MD, Reyes RM, Limongi JE. Natural infection of Lutzomyia ovallesi (Diptera: Psychodidae) with parasites of the Leishmania braziliensis complex in a restricted focus os cutaneous leishmaniasis in Northern Venezuela. Mem Inst Oswaldo Cruz. 1988; 83(3):393–4. https://doi.org/10.1590/s0074-02761988000300019 PMID: 3271937
- Azevedo ACR, Rangel EF, Costa EM, David J, Vasconcelos AW, Lopes UG. Natural infection of Lutzomyia (Nyssomyia) whitmani (Antunes & Coutinho, 1939) by Leishmania of the braziliensis complex in Baturite, Ceará state, Northeast Brazil. Mem Inst Oswaldo Cruz. 1990; 85:1.
- Bonfante-Garrido R, Urdaneta R, Urdaneta I, Alvarado J. Natural Infection of Lutzomyia trinidadensis (Diptera: Psychodidae) with Leishmania in Barquisimeto, Venezuela. Mem Inst Oswaldo Cruz. 1990; 85(4):477. https://doi.org/10.1590/s0074-02761990000400015 PMID: 2152445
- Carvalho GML, Filho JDA, Falcão AL, Rocha Lima ACVM, Gontijo CMF. Naturally Infected Lutzomyia Sand Flies in a Leishmania -Endemic Area of Brazil. Vector-Borne Zoonotic Dis. 2008 Jun; 8(3):407– 14. https://doi.org/10.1089/vbz.2007.0180 PMID: 18429695

- Ashford RW, Desjeux P, DeRaadt P. Estimation of population at risk of infection and number of cases of Leishmaniasis. Parasitol Today. 1992 Mar; 8(3):104–5. PMID: 15463585
- Bezerra-Vasconcelos DR, Melo LM, Albuquerque ÉS, Luciano MCS, Bevilaqua CML. Real-time PCR to assess the Leishmania load in Lutzomyia longipalpis sand flies: Screening of target genes and assessment of quantitative methods. Exp Parasitol. 2011 Nov; 129(3):234–9. <u>https://doi.org/10.1016/j.</u> exppara.2011.08.010 PMID: 21864530
- Nzelu CO, Gomez EA, Cáceres AG, Sakurai T, Martini-Robles L, Uezato H, et al. Development of a loop-mediated isothermal amplification method for rapid mass-screening of sand flies for Leishmania infection. Acta Trop. 2014 Apr; 132:1–6. <u>https://doi.org/10.1016/j.actatropica.2013.12.016</u> PMID: 24388795
- 32. Aransay AM, Scoulica E, Tselentis Y. Detection and Identification of Leishmania DNA within Naturally Infected Sand Flies by Seminested PCR on Minicircle Kinetoplastic DNA. Appl Environ Microbiol. 2000 May 1; 66(5):1933–8. https://doi.org/10.1128/aem.66.5.1933-1938.2000 PMID: 10788363
- 33. de Pita-Pereira D, Cardoso MAB, Alves CR, Brazil RP, Britto C. Detection of natural infection in Lutzomyia cruzi and Lutzomyia forattinii (Diptera: Psychodidae: Phlebotominae) by Leishmania infantum chagasi in an endemic area of visceral leishmaniasis in Brazil using a PCR multiplex assay. Acta Trop. 2008; 107(1):66–9. https://doi.org/10.1016/j.actatropica.2008.04.015 PMID: 18502392
- Marcondes CB, Bittencourt IA, Stoco PH, Eger I, Grisard EC, Steindel M. Natural infection of Nyssomyia neivai (Pinto, 1926) (Diptera: Psychodidae, Phlebotominae) by Leishmania (Viannia) spp. in Brazil. Trans R Soc Trop Med Hyg. 2009 Nov; 103(11):1093–7. https://doi.org/10.1016/j.trstmh.2008.12. 006 PMID: 19178921
- Saraiva L, Carvalho GML, Gontijo CMF, Quaresma PF, Lima ACVMR, Falcão AL, et al. Natural Infection of Lutzomyia neivai and Lutzomyia sallesi (Diptera: Psychodidae) by Leishmania infantum chagasi in Brazil. J Med Entomol. 2009 Sep 1; 46(5):1159–63. <u>https://doi.org/10.1603/033.046.0525</u> PMID: 19769049
- Es-Sette N, Ajaoud M, Bichaud L, Hamdi S, Mellouki F, Charrel RN, et al. Phlebotomus sergenti a common vector of Leishmania tropica and Toscana virus in Morocco. J Vector Borne Dis. 2014 Jun; 51 (2):86–90. PMID: 24947214
- Zorrilla V, De Los Santos MB, Espada L, Santos R del P, Fernandez R, Urquia A, et al. Distribution and identification of sand flies naturally infected with Leishmania from the Southeastern Peruvian Amazon. McDowell MA, editor. PLoS Negl Trop Dis. 2017 Nov 6; 11(11):e0006029 https://doi.org/10.1371/ journal.pntd.0006029 PMID: 29107954
- Ministério da Saúde/SVS—Sistema de Informação de Agravos de Notificação—Sinan Net < <a href="http://tabnet.datasus.gov.br/cgi/deftohtm.exe?sinannet/cnv/leishvba.def">http://tabnet.datasus.gov.br/cgi/deftohtm.exe?sinannet/cnv/leishvba.def</a>>
- Young D, Duncan M. Guide to the indentification and geographic distribution of Lutzomyia sand flies in Mexico, the west Indies, central and south America (Diptera: Psychodidae). Mem Am Entomol Inst. 1994 Jun 19; 54.
- 40. Langeron, M. Precis de microscopie. v. 7 ed. Paris: Masson e Cie, p. 1429, 1949.
- Bauzer LGSR Souza NA, Ward RD Kyriacou CP, Peixoto AA. The period gene and genetic differentiation between three Brazilian populations of Lutzomyia longipalpis. Insect Mol Biol. 2002 Aug; 11 (4):315–23. PMID: 12144696
- Francino O, Altet L, Sánchez-Robert E, Rodriguez A, Solano-Gallego L, Alberola J, et al. Advantages of real-time PCR assay for diagnosis and monitoring of canine leishmaniosis. Vet Parasitol. 2006 Apr; 137 (3–4):214–21. https://doi.org/10.1016/j.vetpar.2006.01.011 PMID: 16473467
- 43. Solcà MDS, Bastos LA, Guedes CES, Bordoni M, Borja LS, Larangeira DF, et al. Evaluating the Accuracy of Molecular Diagnostic Testing for Canine Visceral Leishmaniasis Using Latent Class Analysis. Traub-Csekö YM, editor. PLoS One. 2014 Jul 30; 9(7):e103635. https://doi.org/10.1371/journal.pone. 0103635 PMID: 25076494
- 44. Borja LS, Sousa OMF de, Solcà M da S, Bastos LA, Bordoni M, Magalhães JT, et al. Parasite load in the blood and skin of dogs naturally infected by Leishmania infantum is correlated with their capacity to infect sand fly vectors. Vet Parasitol. 2016 Oct; 229:110–7. <u>https://doi.org/10.1016/j.vetpar.2016.10</u>. 004 PMID: 27809965
- 45. Paiva BR de, Secundino NFC, Pimenta PFP, Galati EAB, Andrade Junior HF, Malafronte R dos S. Padronização de condições para detecção de DNA de Leishmania spp. em flebotomíneos (Diptera, Psychodidae) pela reação em cadeia da polimerase. Cad Saude Publica. 2007 Jan; 23(1):87–94. PMID: 17187107
- 46. De Souza CM, Pessanha JE, Barata RA, Monteiro ÉM, Costa DC, Dias ES. Study on phlebotomine sand fly (Diptera: Psychodidae) fauna in Belo Horizonte, state of Minas Gerais, Brazil. Mem Inst Oswaldo Cruz. 2004; 99(8):795–803. <u>https://doi.org/10.1590/s0074-02762004000800003</u> PMID: 15761593

- 47. Oliveira DMS de, Saraiva EM, Ishikawa EAY, Sousa AAA de, Silva EO da, Silva IM da. Distribution of phlebotomine fauna (Diptera: Psychodidae) across an urban-rural gradient in an area of endemic visceral leishmaniasis in northern Brazil. Mem Inst Oswaldo Cruz. 2011 Dec; 106(8):1039–44. https://doi.org/10.1590/s0074-02762011000800023 PMID: 22241130
- 48. Jeraldo V de LS, Góes MA de O, Casanova C, Melo CM de, Araújo ED de, Filho SPB, et al. Sand fly fauna in an area endemic for visceral leishmaniasis in Aracaju, State of Sergipe, Northeast Brazil. Rev Soc Bras Med Trop. 2012; 45(3):318–22. PMID: 22760129
- Werneck GL. Forum: geographic spread and urbanization of visceral leishmaniasis in Brazil. Introduction. Cad Saude Publica [Internet]. 2008; 24(12):2937–40. PMID: 19082285
- 50. Michalsky ÉM, de Sena Guedes K, de Oliveira Lara Silva F, França-Silva JC, Dias CLF, Barata RA, et al. Infecção natural de Lutzomyia (Lutzomyia) longipalpis (Diptera: Psychodidae) por Leishmania infantum chagasi em flebotomíneos capturados no município de Janaúba, estado de Minas Gerais, Brasil. Rev Soc Bras Med Trop. 2011; 44(1):58–62. PMID: 21340410
- de Carvalho MR, Valença HF, da Silva FJ, de Pita-Pereira D, de Araújo Pereira T, Britto C, et al. Natural Leishmania infantum infection in Migonemyia migonei (França, 1920) (Diptera:Psychodidae:Phlebotominae) the putative vector of visceral leishmaniasis in Pernambuco State, Brazil. Acta Trop. 2010 Oct; 116(1):108–10. https://doi.org/10.1016/j.actatropica.2010.03.009 PMID: 20457120
- Rosa J, Pereira DP, Brazil RP, Filho JDA, Salomón O, Szelag E. Natural infection of cortelezzii complex (Diptera: Psychodidae: Phlebotominae) with Leishmania braziliensis in Chaco, Argentina. Acta Trop. 2012 Aug; 123(2):128–31. https://doi.org/10.1016/j.actatropica.2012.04.008 PMID: 22569560
- **53.** Casaril A, Monaco NZ, de Oliveira E, Eguchi G, Filho AC, Pereira L, et al. Spatiotemporal analysis of sand fly fauna (Diptera: Psychodidae) in an endemic area of visceral leishmaniasis at Pantanal, central South America. Parasit Vectors. 2014; 7(1):364.
- 54. Oliveira EF de, Casaril AE, Fernandes WS, Ravanelli M de S, Medeiros MJ de, Gamarra RM, et al. Monthly Distribution of Phlebotomine Sand Flies, and Biotic and Abiotic Factors Related to Their Abundance, in an Urban Area to Which Visceral Leishmaniasis Is Endemic in Corumbá, Brazil. Lorenzo MG, editor. PLoS One. 2016 Oct 26; 11(10):e0165155. https://doi.org/10.1371/journal.pone.0165155 PMID: 27783667
- 55. Miranda JC, Reis E, Schriefer A, Gonçalves M, Reis MG, Carvalho L, et al. Frequency of Infection of Lutzomyia Phlebotomines with Leishmania braziliensis in a Brazilian Endemic Area as Assessed by Pinpoint Capture and Polymerase Chain Reaction. Mem Inst Oswaldo Cruz. 2002 Mar; 97(2):185–8. https://doi.org/10.1590/s0074-02762002000200006 PMID: 12016439
- 56. Missawa NA, Michalsky ÉM, Fortes-Dias CL, Santos Dias E. Lutzomyia longipalpis naturally infected by Leishmania (L.) chagasi in Várzea Grande, Mato Grosso State, Brazil, an area of intense transmission of visceral leishmaniasis. Cad Saude Publica. 2010 Dec; 26(12):2414–9. PMID: 21243236
- Paiva BR, Oliveira AG, Dorval MEMC, Galati EAB, Malafronte RS. Species-specific identification of Leishmania in naturally infected sand flies captured in Mato Grosso do Sul State, Brazil. Acta Trop. 2010 Jul; 115(1–2):126–30. https://doi.org/10.1016/j.actatropica.2010.02.013 PMID: 20219438
- Rocha LS, Falqueto A, dos Santos CB, Ferreira AL, da Graça GC, Grimaldi G, et al. Survey of natural infection by Leishmania in sand fly species collected in southeastern Brazil. Trans R Soc Trop Med Hyg. 2010 Jul; 104(7):461–6. https://doi.org/10.1016/j.trstmh.2010.02.005 PMID: 20346478
- 59. Pita-Pereira D de, Souza GD, Pereira T de A, Zwetsch A, Britto C, Rangel EF. Lutzomyia (Pintomyia) fischeri (Diptera: Psychodidae: Phlebotominae), a probable vector of American Cutaneous Leishmania-sis: Detection of natural infection by Leishmania (Viannia) DNA in specimens from the municipality of Porto Alegre (RS), Brazil, using. Acta Trop. 2011 Dec; 120(3):273–5. https://doi.org/10.1016/j. actatropica.2011.09.004 PMID: 21939631
- Cunha RC, Andreotti R, Cominetti MC, Silva EA. Detection of Leishmania infantum in Lutzomyia longipalpis captured in Campo Grande, MS. Rev Bras Parasitol Veterinária. 2014 Jun; 23(2):269–73.
- Brighente KB dos S, Cutolo AA, Motoie G, da Silva Meira-Strejevitch C, Pereira-Chioccola VL. Molecular detection of Leishmania (Leishmania) infantum in phlebotomine sand flies from a visceral leishmaniasis endemic area in northwestern of São Paulo State, Brazil. Acta Trop. 2018 May; 181(October 2017):1–5. https://doi.org/10.1016/j.actatropica.2018.01.012 PMID: 29373820
- Paiva BR, Secundino NFC, Nascimento JC, Pimenta PFP, Galati EAB, Junior HFA, et al. Detection and identification of Leishmania species in field-captured phlebotomine sand flies based on mini-exon gene PCR. Acta Trop. 2006 Oct; 99(2–3):252–9. <u>https://doi.org/10.1016/j.actatropica.2006.08.009</u> PMID: 17055444
- Silva EA, Andreotti R, Dias ES, Barros JC, Brazuna JCM. Detection of Leishmania DNA in phlebotomines captured in Campo Grande, Mato Grosso do Sul, Brazil. Exp Parasitol. 2008 Jul; 119(3):343–8. https://doi.org/10.1016/j.exppara.2008.03.011 PMID: 18456262

- 64. Pita-Pereira D de, Lis R, Oliveira MP, Lima RB, Pereira BA, Moreira OC, et al. SYBR Green-based Real-Time PCR targeting kinetoplast DNA can be used to discriminate between the main etiologic agents of Brazilian cutaneous and visceral leishmaniases. Parasites and Vectors. 2012; 5(1):1–9. https://doi.org/10.1186/1756-3305-5-1
- 65. Ferreira TS, Timbó RV, Minuzzi-Souza TTC, de Almeida Rocha D, Neiva M, de Albuquerque Ribeiro J, et al. High molecular prevalence of Leishmania in phlebotomine sand flies fed on chicken blood in Brazil. Vet Parasitol. 2018 Aug; 259:80–4. https://doi.org/10.1016/j.vetpar.2018.07.004 PMID: 30056989
- Resende MC, Camargo MCV, Vieira JRM, Nobi RCA, Porto NMN, Oliveira CDL, et al. Seasonal variation of Lutzomyia longipalpis in Belo Horizonte, State of Minas Gerais. Rev Soc Bras Med Trop. 2006; 39(1):51–5. https://doi.org//S0037-86822006000100010 PMID: 16501767
- **67.** Mestre GL da C, Ribeiro ALM, Miyazaki RD, Rodrigues JS V, Almeida A do BPF de, Sousa VRF, et al. Phlebotomine sand flies and canine infection in areas of human visceral leishmaniasis, Cuiabá, Mato Grosso. Rev Bras Parasitol Veterinária. 2011; 20:228–34.
- Pinto I de S, Ferreira AL, Valim V, dos Santos Carvalho F, da Silva GM, Falcão AL, et al. Sand fly vectors (Diptera, Psychodidae) of American visceral leishmaniasis areas in the Atlantic Forest, State of Espírito Santo, southeastern Brazil. J Vector Ecol. 2012 Jun; 37(1):90–6. https://doi.org/10.1111/j. 1948-7134.2012.00204.x PMID: 22548541
- 69. Rangel O, Sampaio SMP, Ciaravolo RM de C, Holcman MM. The distribution pattern of Lutzomyia longipalpis (Diptera: Psychodidae) in the peridomiciles of a sector with canine and human visceral leishmaniasis transmission in the municipality of Dracena, São Paulo, Brazil. Mem Inst Oswaldo Cruz. 2012 Mar; 107(2):163–9. https://doi.org/10.1590/s0074-02762012000200003 PMID: 22415253
- 70. Barata RA, Silva JCF da, Costa RT da, Fortes-Dias CL, Silva JC da, Paula EV de, et al. Phlebotomine sand flies in Porteirinha, an area of American visceral leishmaniasis transmission in the State of Minas Gerais, Brazil. Mem Inst Oswaldo Cruz. 2004 Aug; 99(5):481–7. <u>https://doi.org/10.1590/s0074-02762004000500004 PMID</u>: 15543410
- França-Silva JC, Barata RA, Costa RT da, Monteiro ÉM, L. Machado-Coelho GL, Vieira EP, et al. Importance of Lutzomyia longipalpis in the dynamics of transmission of canine visceral leishmaniasis in the endemic area of Porteirinha Municipality, Minas Gerais, Brazil. Vet Parasitol. 2005 Aug; 131(3– 4):213–20. https://doi.org/10.1016/j.vetpar.2005.05.006 PMID: 15975718
- 72. Gonçalves R, Soares DC, Guimarães RJ de PS e, Santos WS, Sousa GCR de, Chagas AP, et al. Diversity and ecology of sand flies (Psychodidae: Phlebotominae): foci of cutaneous leishmaniasis in Amazon Region, Brazil. Rev Pan-Amazônica Saúde. 2016 Dec; 7:133–42.
- 73. Fernandes W de S, Borges LM, Casaril AE, Oliveira EF de, Infran J de OM, Piranda EM, et al. Sand fly fauna (Diptera: Psychodidae) in an urban area, Central-West of Brazil. Rev Inst Med Trop Sao Paulo. 2017 Aug 24; 59:1–8.
- 74. Oliveira AM, López RVM, Dibo MR, Rodas LAC, Guirado MM, Chiaravalloti-Neto F. Dispersion of Lutzomyia longipalpis and expansion of visceral leishmaniasis in São Paulo State, Brazil: identification of associated factors through survival analysis. Parasit Vectors. 2018 Dec 10; 11(1):503. https://doi.org/ 10.1186/s13071-018-3084-1 PMID: 30201037
- 75. Dias-Lima AG, Guedes MLS, Sherlock IA. Horizontal stratification of the sand fly fauna (Diptera: Psychodidae) in a transitional vegetation between caatinga and tropical rain forest, state of Bahia, Brazil. Mem Inst Oswaldo Cruz. 2003 Sep; 98(6):733–7. <u>https://doi.org/10.1590/s0074-02762003000600004</u> PMID: 14595447
- 76. Nieto P, Malone JB, Bavia ME. Ecological niche modeling for visceral leishmaniasis in the state of Bahia, Brazil, using genetic algorithm for rule-set prediction and growing degree day-water budget analysis. Geospat Health. 2006 Nov 1; 1(1):115. https://doi.org/10.4081/gh.2006.286 PMID: 18686237
- 77. Pinto I de S, Ferreira AL, Valim V, dos Santos Carvalho F, da Silva GM, Falcão AL, et al. Sand fly vectors (Diptera, Psychodidae) of American visceral leishmaniasis areas in the Atlantic Forest, State of Espírito Santo, southeastern Brazil. J Vector Ecol. 2012 Jun; 37(1):90–6. https://doi.org/10.1111/j. 1948-7134.2012.00204.x PMID: 22548541
- 78. Dias ES, França-Silva JC, Da Silva JC, Monteiro ÉM, Paula KM De, Macedo C, et al. Flebotomíneos (Diptera: Psychodidae) de um foco de leishmaniose tegumentar no Estado de Minas Gerais Sand flies | (Diptera: Psychodidae) in an outbreak of cutaneous leishmaniasis in the State of Minas Gerais. Rev Soc Bras Med Trop. 2007; 40(1):49–52.
- 79. Ximenes M de FF de M, Castellón EG, De Souza M de F, Menezes AAL, Queiroz JW, Macedo e Silva VP, et al. Effect of abiotic factors on seasonal population dynamics of Lutzomyia longipalpis (Diptera: Psychodidae) in northeastern Brazil. J Med Entomol. 2006; 43(5):990–5. PMID: 17017238
- 80. Amóra SSA, Bevilaqua CML, Dias E de C, Feijó FMC, Oliveira PGM de, Peixoto GCX, et al. Monitoring of Lutzomyia longipalpis Lutz & Neiva, 1912 in an area of intense transmission of visceral leishmaniasis in Rio Grande do Norte, Northeast Brazil. Rev Bras Parasitol Veterinária. 2010 Mar; 19(1):39–43.