Sección Médica

Morphometric analysis of *Chagasia fajardi* (Diptera: Culicidae) populations in Brazil

Análisis morfométrico de poblaciones de Chagasia fajardi (Diptera: Culicidae) del Brasil

JÚLIA DOS SANTOS SILVA¹, ANA LAURA CARBAJAL DE LA FUENTE^{2,3}, NICOLAU MAUÉS SERRA-FREIRE⁴, ANTHONY ÉRICO GUIMARÃES⁵, JULIANA SOARES SARMENTO⁶ and JERONIMO ALENCAR⁵

Abstract: The objective of the present study was to evaluate morphological variability in four populations of *Chagasia fajardi* using classic morphometry. Seven variables of the body were measured in specimens of *Ch. fajardi* from four biogeographical regions of Brazil, including three populations from the Atlantic Forest biome and one population from the Cerrado biome, were measured. Using analysis of variance (ANOVA), the diagnostic characteristic of the black band on the hindtarsus (Ta-III₃) presented strong potential for demonstrating dissimilarity between populations, which were all different from each other. This was followed by the white band on the hindtarsus (Ta-III₅) and the wing length (Wl), which showed the presence of two populations similar between themselves, but different from the others. However, multivariate analysis and UPGMA dendograms showed close proximity between populations 2 and 4 and between populations 1 and 3. Morphological variables of *Ch. fajardi* populations from different regions of Brazil are discussed

Key words: Classic morphometry. ANOVA. Multivariate analyses. Mosquitoes.

Resumen: El presente estudio tuvo como objetivo evaluar la variabilidad morfológica en cuatro poblaciones de *Chagasia fajardi* utilizando morfometría clásica. Se midieron siete variables del cuerpo de los especímenes de cuatro regiones biogeográficas brasileñas, incluyendo tres poblaciones provenientes de los biomas "Mata" Atlántica y una población del Cerrado. Usando el análisis de varianza (ANOVA), el carácter diagnóstico banda negra del tarsómero posterior tres (TA-III₃) mostró un gran potencial para determinar las diferencias entre poblaciones, las cuales fueron todas distintas unas de las otras. Esto se siguió por la banda blanca del tarso cinco posterior (Ta-III₅) y la longitud del ala (WI), que mostraron la presencia de dos poblaciones similares entre sí pero diferentes de las otras poblaciones, siendo estas últimas también diferentes entre sí. Sin embargo, el análisis multivariado y dendrograma UPGMA mostraron la gran proximidad entre las poblaciones 2 y 4, y entre las poblaciones 1 y 3. Se discuten las variables morfológicas de poblaciones pertenecientes a *Ch. fajardi* de diferentes regiones brasileñas.

Palabras clave: Morfometría clásica. ANOVA. Análisis multivariado. Mosquitos.

Introduction

According to Harbach (2007), the subfamily Anophelinae includes 476 formally described species. The traditional classification of this subfamily includes three genera: Anopheles, Bironella and Chagasia. The genus Chagasia Cruz is comprised of five species (Harbach and Howard 2009). The mosquitoes of this genus have morphological characteristics similar to those of the subfamily Culicinae, but are considered to be primitive because of the presence of eight chromosomes (2n = 8), as identified in *Chagasia bathana* (Dyar, 1928) (Kreutzer 1978). In the other Culicidae, the number of chromosomes is 2n = 6. According to Forattini (2002), the presence of multiple floaters in the eggs is indicative of the existence of an evolutionary process associated with the genus Chagasia, in which an ancestral characteristic from a single pair of chromosomes underwent adaptation, as observed in the anophelines.

Baerg & Boreham (1974) demonstrated experimentally that females of Ch. bathana can fed using human blood. There was no evidence of any degree of endophilia, and they were not incriminated as vectors of any pathogenic agent for human beings or domesticated animals (Forattini 2002). These mosquitoes have typically sylvatic behavior and are found in forested environments. They appear to be strictly zoophilic with acrodendrophile tendencies (Galindo and Trapido 1957). Their distribution is restricted to the Neotropical region, and they are usually found in mountainous areas (Causey et al. 1945). The latest compilation of the geographical distribution of Chagasia was presented by Harbach and Howard (2009) and comprises Ch. ablusa Harbach, 2009 (Colombia and Peru); Ch. bonneae Root, 1927 (Bolivia, Brazil, Colombia, Ecuador, French Guiana, Guyana, Peru, Suriname and Venezuela); Ch. fajardi (Lutz, 1904) (Argentina, Bolivia, Brazil, Colombia and Guyana); Ch. rozeboomi Causey, Deane and Deane, 1944 (Brazil); and Ch. bathana (Belize, Colombia,

¹ Ph. D. Department of Entomology, National Museum, Federal University of Rio de Janeiro, Brazil, ss_juliass@yahoo.com.br. ² Ph. D. Leishmaniasis Transmitter Laboratory, Oswaldo Cruz Institute/Fiocruz, Av. Brasil 4365, CEP: 21040-360, Manguinhos, Rio de Janeiro, Brazil, analauracf@ege.fcen.uba.ar. ³ Departament Ecology, Genetic and Evolution, Eco-Epidemiology Laboratory, Faculty of Exact and Natural Sciences, University of Buenos Aires, Argentina. ⁴ Ph. D. Ixoxides Laboratory and National Reference Center for Rickettsial Vectors, Oswaldo Cruz Institute/Fiocruz, Av. Brasil 4365, CEP: 21040-360, Manguinhos, Rio de Janeiro, Brazil, nnsf@ioc.fiocruz.br. ⁵ Diptera Laboratory, Oswaldo Cruz Institute/Fiocruz, Av. Brasil 4365, CEP: 21040-360, Manguinhos, Rio de Janeiro, Brazil, anthony@ioc.fiocruz.br, jalencar@ioc.fiocruz.br, corresponding author. ⁶ M. Sc. Graduation Program in Animal biology, Rural Federal University of Rio de Janeiro, Brasil.

Costa Rica, Ecuador, Honduras, French Guiana, Guatemala, Mexico, Nicaragua, Panama and Venezuela).

Morphometric analysis is a useful technique for solving taxonomic problems, such as studying and delimiting populations of the same species, or recognizing cryptic species in sympatric or allopatric populations (Calle et al. 2002). It has also been shown to be useful for correlations with other phylogenetic techniques, such as isoenzyme analysis (Dujardin et al. 2000), and for differentiation of insect populations of medical importance (Dujardin et al. 1997; Dujardin and Slice 2006; Dujardin 2008). There are few papers which show the use of morphometry in Culicidae (Jirakanjanakit et al. 2008; Suman et al. 2009; Morais et al. 2010; Kanojia et al. 2010). Some authors used morphometry successfully to identify some species of the subgenus Anopheles Nyssorhynchus from Colombia and Venezuela, including Delgado and Rubio-Palis (1993), Rubio-Palis (1998, 2000), Calle et al. (2002) and Laurito et al. (2008).

In order to assess the phenotypic variability of *Ch. faja-rdi*, a comparative study of body size in four Brazilian populations of this species was performed.

Materials and Methods

Insect samples. The four Brazilian female populations used for this study came from the following locations: 1) Bocaina National Park, Municipality of São José do Barreiro, State of São Paulo, between 22°45'S 44°37' - 44°43'W, which is in the Atlantic Forest biome at an altitude between 1495 and 1642 masl; 2) Corumbá Hydroelectric Plant, Municipality of Caldas Novas, State of Goiás, 17°45'S 48°30'W, which is in the savanna (Cerrado) biome at an altitude of 686 masl; 3) Itatiaia National Park, Municipality of Itatiaia, State of Rio de Janeiro, 22°29'S 44°33'W, in the Atlantic Forest at an altitude of 610 masl; and 4) Iguaçu National Park, Municipality of Foz do Iguaçu, State of Paraná, between 25° 37- 25°40'S



Figure 1. Locations from Brazil where the mosquitoes analyzed originated.

54°26′- 54°28′W, in the Atlantic Forest at an altitude between 180 and 222 masl. (Fig. 1). The females were collected using a manual suction tube (Castro collector) and transferred to plastic bottles with a height 5 cm and diameter 3 cm which contained a layer of cotton wool soaked in paraformol at the bottom, overlain with filter paper. The specimens were taken to the laboratory on the same day.

Mosquitoes populations examined were from ecological and epidemiological studies developed by the staff of *Laboratório de Diptera do Instituto Oswaldo Cruz - Fiocruz*. Populations were collected during the following sampling periods: Bocaina National Park, January 1991 to December 1992; Corumbá Hydroelectric Plant, January 1996 to January 1997; Itatiaia National Park, August 2002 to July 2003; and Iguaçu National Park, January 1995 to November 1996.

The mosquitoes were identified at the laboratory using the adult identification keys produced by Causey *et al.* (1945), Forattini (2002) and Harbach and Howard (2009). The genus and subgenus abbreviations followed the system proposed by Reinert (2001) and the terminology for description used was in accordance with Harbach and Knight (1980). Specimens for reference were deposited in the Entomological Collection of the Oswaldo Cruz Institute (IOC), within the Atlantic Forest Collection and the Cerrado Collection.

Measured characteristics. A total of 119 females from the field were examined, including 30 specimens from each locality (except for population 3 with n = 29), and seven variables on the mosquito specimens were measured: 1) the black band on the hindtarsus (Ta-III₅); 2) the white band on the hindtarsus (Ta-III₅); 3) the black band on the hindtarsus (Ta-III₃); 5) the total length of the hindtarsus (Ta-III); 6) the wing length (Wl); and 7) the proboscis length (P) (Fig. 2). Measurements on these mosquito characteristics were made using a stereoscopic microscope (ZEISS Stemi SV6®) with a 10 X eyepiece and 43 X. A micrometer (100 divisions = 10 mm) was calibrated using a micrometric lamina (0.001 mm).

Statistical analysis. The morphometric data were quantitatively analyzed, obtaining the means and standard deviations.

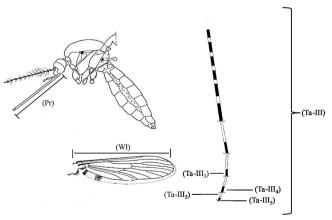


Figure 2. Variables analyzed of *Chagasia fajardi*: 1) the black band on the hindtarsus (Ta-III₅); 2) the white band on the hindtarsus (Ta-III₅); 3) the black band on the hindtarsus (Ta-III₄); 4) the black band on the hindtarsus (Ta-III₃); 5) the total length of the hindtarsus (Ta-III); 6) the wing length (WI); and 7) the proboscis length (Pr).

In order to check the precision of the data, the precision test (P) was used. The calculation was made from pairs of measurements on 30% of the specimens, which were selected randomly. In order to exclude doubtful variables, the reliability of the data was retested by means of the exactness method (E). The exactness was calculated between the maximum and minimum values of each variable, multiplied by one hundred.

The variance of the population was estimated using single-factor analysis of variance (ANOVA). The estimate of the variance was calculated by combining the variances between the samples relating to the modes. The "F" ratio was calculated to indicate how different modes related to the variable in each population studied. The significance of the minimum amplitude was analyzed using the Duncan test. This ANOVA and the "F" region were used to test the hypothesis that the effects from the parameters of the seven diagnostic characteristics were real. The relationships between the central trend measurements proved that all the samples of the four populations had normal distribution and similar variances, and this made it possible to test the variance relating to the mode. The specific differences between the means of the samples coming from the four regions investigated were tested, two at a time, using the "t" test to investigate significance between pairs of mean samples values, with a cutoff of a = 10%.

Variables with significant differences were used for a discriminant analysis (Sokal and Sneath 1963; Morrison 1976) between populations. The data from each population were log-transformed to obtain functions that enabled better identification of the study groups. For population analysis, Mahalanobis distances and their statistical significance were calculated using permutation tests (1000 runs each) after Bonferroni correction. Cluster analysis based on Mahalanobis distances was used to build an UPGMA (unweighted pair-group method with arithmetic average) dendogram. The analyses were performed using the *PADwin* software, version 81a (Dujardin 2003).

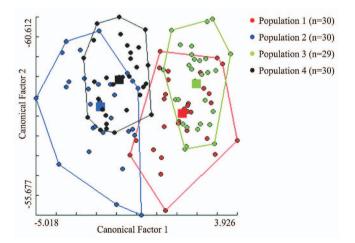


Figure 3. Canonical variate analysis on four *Chagasia fajardi* populations based on five variables of the mosquito body, showing centroid distribution (multivariate mean) of canonical factors 1 (CF1) and 2 (CF2). The polygon represents the dispersion of points corresponding to each population. Canonical factors 1 (CF1) explained 95% of the variance, while CF2 explained 0.05%.

Results

The means and standard deviations of the descriptive statistics on the seven measured variables in each population are presented in Table 1, with the graduation unit values already converted into millimeters. From these results, population 1 was observed to have the lowest values for the seven variables measured.

ANOVA revealed the diagnostic characteristic of the black band on the hindtarsus (Ta-III₃) had strong potential to show dissimilarity between the populations. This was followed by the white band on the hindtarsus (Ta-III₅) and the wing length (Wl), which showed the two populations ware similar and different from the others, which also differed from each other. The variable black band on the hindtarsus (Ta-III₄) showed the greatest similarity among the populations. Using groups of two and three populations, for different sets of variable studied in the populations, it had differences or similarities with regard to the different sets of the tested characteristics (Table 2).

Through discriminant analysis, canonical factor 1 (CF1) separated populations 2 and 4 from populations 1 and 3 (Fig. 3). Canonical Factor 1(CF1) explained 95% of the variance while CF2 explained 0.05%. The Mahalanobis distances between the four populations were highly significant (P < 0.001) (Table 3). These distances were used in UPGMA (unweighted pair-group method with arithmetic average) cluster analyses to produce dendrogram. The derived UPGMA dendrogram showed two clearly separated clusters formed by populations 1 and 3 in one position and populations 2 and 4 in another position (Fig 4). After 1000 permutations, 63% of population 1 specimens were correctly reclassified, while population 2, 3 and 4 specimens had reclassification rates of 70%, 68% and 63% respectively (Table 4).

Discussion

According to Harbach and Howard (2009), *Chagasia* is a genus with few taxa that forms a homogenous set of species with few morphological distinctions. Molecular data would probably be necessary to elucidate the phylogenetic relationships within the genus. *Chagasia ablusa* has been erroneously identified as *Ch. fajardi*, based on the superficial morphological similarities of the adults (Harbach and Howard 2009; Berti *et al.* 2011). These similarities indicate that *Ch. ablusa* is morphologically closer to *Ch. fajardi* than any other spe-

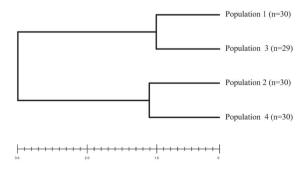


Figure 4. UPGMA dendogram derived from Mahalanobis distances based on five variables of the body in the *Chagasia fajardi* populations.

^{* 1:} Bocaina Range National Park; 2: Corumbá Hydroelectric Plant, Goiás; 3: Itatiaia National Park; 4: Iguaçu National Park.

Fable 1. Mean, standard deviation (SD) and coefficient of variation (CV) of the four populations (Pop.) of *Chagasia fajardi* (Diptera: Culicidae).

																				,	
Pop.		Ta-IIIs black band		e"	Ta-IIIs white band		1	Ta-III.4 black band			Ta-III3 black band		7	Ta-III total lenght		*	WI wing length			P P. length	
	Mean	SD	CV	Mean	SD	CV	Mean	SD	CV	Mean	SD	CV	Mean	S	CV	Mean	SD	CV	Mean	SD	CV
-	0.101	0.022	22.054	0.203	0.024	11.661	0.122	0.018	14.875	0.218	0.038	17.302	5.190	0.301	5.808	3.381	0.168	4.960	1.635	0.091	5.548
2	0.077	0.015	19.150	0.186	0.018	9.677	0.131	0.015	11.587	0.175	0.023	13.067	4.636	0.271	5.855	2.937	0.180	6.137	1.403	0.269	19.192
33	0.105	0.024	22.960	0.221	0.028	12.765	0.135	0.029	21.669	0.244	0.041	16.852	5.302	0.313	5.904	3.500	0.144	4.107	1.673	0.065	3.899
4	0.081	0.018	21.933	0.199	0.020	10.194	0.135	0.017	12.472	0.195	0.023	12.023	4.719	0.213	4.522	3.025	0.137	4.535	1.455	0.072	4.918

on the hindtarsus; Ta-III; the black band on the hindtarsus; Ta-III; the black band on the hindtarsus; Ta-III; total length of the posterior tarsi; WI: wing length; and Corumbá Hydroelectric Plant, Goiás; Pop. 3: Itatiaia National Park; Pop. 4: Iguaçu National Park. * $Ta-III_s$: the black band on the hindtarsus; $Ta-III_s$: the white band proboscis length. Pop. 1: Bocaina Range National Park; Pop. 2: cies of *Chagasia*, which is a debatable matter (Harbach and Howard 2009). The structure of the male genitalia and the dark scales in the central area, dorsal face, and distal half of the wings suggest that *Ch. ablusa*, *Ch. fajardi* and *Ch. roze-boomi* are relatively closer to each other than to *Ch. bathana* and *Ch. bonneae* (Harbach and Howard 2009). Although *Ch. fajardi* is considered to be a species without epidemiological importance for disease transmission, and has essentially zoophilic habits, its feeding habits have been observed to be highly eclectic and opportunistic, such that it reacted to all the antiserums analyzed by Alencar *et al.* (2005) and Silva (2011). Moreover, specimens that were positive for human/primate antiserum were found in all the geographical regions studied (Alencar *et al.* 2005).

In comparison with DNA techniques, morphometry is considered to be an inexpensive tool with rapid results. It has made important contributions towards medical entomology (Dujardin and Slice 2006) and is able to make use of specimens in entomological collections without damaging them (Avtekin et al. 2007). The results showed by Mahalanobis distances suggest morphometric variability exists among populations of Ch. fajardi, thereby showing groupings between populations 1 and 3 and between populations 2 and 4. The discriminant analysis provided satisfactory individual reclassifiction rates as showed by Bouyer et al. (2007) for Diptera (Glossinidae). The phenotypic variability encountered in the populations analyzed may have been influenced in different ecological levels, with the environment being able to influence the rate of reproduction by speeding up or reducing the setting of a happened morphologic modification. The four populations of Ch. fajardi come from four Brazilian states and two different biomes. The population 1 and 3 are geographically closer and correspond to the biome of Mata Atlântica. Population 2 and 4 are from the state of Goiás and Paraná respectively and belong to the *cerrado* biome. Bidou & Marti (2008) described the geographic and climatic factors related to a body-size cline in insects (Orthoptera: Acrididae). They showed that the variability of body size increases with latitude and altitude in males and females. This could be a hypothesis for morphological variability of *Chagasia* populations coming from distinct geographical areas. According to Py-Daniel and Fernandes (2005), ecological differentiation may lead to morphological and behavioral differences between the sexes, such as specialization of males and females regarding their intake of different types of foods. However, sexual selection mechanisms are the factors leading to the appearance and fixation of dimorphic sexual characteristics, through females' choice of males and competition between males, or through males' choice of females and competition between females (Py-Daniel and Fernandes, 2005).

Based on ANOVA, morphological differentiation of *Ch. fajardi* has been found in allopatric populations, without a defined pattern of modification. The diagnostic characteristic of the black band on the hindtarsus (Ta-III₃) is the variable that best defines the differentiation. Causey, Deane and Deane (1945) reported that the species *Ch. fajardi* and *Ch. bathana* presented similarity regarding the Ta-III₅, with black marking on 1/3 to 1/4 of their extent. According to Harbach and Howard (2009), posterior tarsal segments 2-5 in adult specimens of *Ch. ablusa* differed from those of *Ch. bathana* in that they did not present a dark postbasal band. In addition, the length of the apical dark band on the hindtarsus Ta-III₂ was different from that of *Ch. fajardi* in 92%.

Table 2. Results from ANOVA, Duncan test and "F" ratio for testing the characteristics or attributes of specimens of *Chagasia fajardi* (Diptera: Culicidae) from four different geographical areas from Brazil.

Population	Mean/Standar Deviation (SD)	Ta-III ₅ black band	Ta-III ₅ white band	Ta-III₄ black band	Ta-III ₃ black band	Ta-III total length	Wl wing length	P P. length
1	Mean	0.101 A	0.203 A	0.122 A	0.218 A	5.190 A	3.381 A	1.673 A
1	SD	0.022	0.024	0.018	0.038	0.301	0.171	0.066
2	Mean	0.077 B	0.186 B	0.131 B	0.175 B	4.636 B	3.381 A	1.635 A
2	SD	0.015	0.018	0.015	0.023	0.276	0.168	0.092
3	Mean	0.105 A	0.221 C	0.135 B	0.244 C	5.300 A	3.500 B	1.399 B
3	SD	0.027	0.029	0.030	0.042	0.318	0.146	0.234
4	Mean	0.081 AC	0.199 A	0.135 B	0.195 D	4.719 B	3.025 C	1.455 B
	SD	0.018	0.021	0.017	0.024	0.024	0.217	0.073

^{*} The same letter in the same column indicate a non significant difference between the means, while different letters in the same column indicate a significant difference between the modes with a 5% chance of error ($\mu = 5.0\%$).

According to Forattini (2002), modifications to populations can occur in two essential manners. In one of them, the population becomes diversified into two or more groups, each one following its own evolution; in the other, the population becomes progressively differentiated as time goes by (Forattini 2002).

The morphometric analysis used in the present study made it possible to show the morphological variability of the populations of *Ch. fajardi* from four different geographical regions of Brazil. The evidence encountered justifies extending the study to search for other variables and tools for achieving as much discrimination between the adult populations as possible.

Table 3. Mahalanobis distances (MD) for the four Brazilian populations (Pop) analyzed of *Chagasia fajardi* (Diptera: Culicidae). Distances were significant (*) at p < 0.00833 after Bonferroni correction; (p) = significant value. Pop 1: Bocaina Range National Park; Pop 2: Corumbá Hydroelectric Plant, Goiás; Pop 3: Itatiaia National Park; Pop 4: Iguaçu National Park.

	MD (p) Pop. 1	MD (p) Pop. 2	MD (p) Pop. 3
Pop. 2	3.65 (0.000)*		
Pop. 3	1.01 (0.052)	4.30 (0.000)*	
Pop. 4	2.94 (0.000)*	1.11 (0.022)	3.44 (0.000)*

Table 4. Reclassification of individuals belonging to four Brazilian populations of *Chagasia fajardi* (Diptera: Culicidae) according to Mahalanobis distances.

Population	N of wing units correctly classified / N wing units	% classified
1	19 / 30	63
2	21 / 30	70
3	20 / 29	68
4	19 / 30	63

N = number

Pop. 1: Bocaina Range National Park; Pop. 2: Corumbá Hydroelectric Plant, Goiás; Pop. 3: Itatiaia National Park; Pop. 4: Iguaçu National Park.

Morphometric analysis is a technique economically favorable and useful for solving taxonomic problems. However, in order to give further support, our results could be corroborated with other techniques, such as molecular biology techniques.

Literature cited

ALENCAR, J.; LOROSA, E. S.; SILVA, J. S.; GIL-SANTANA, H. R.; GUIMARÃES, A. E. 2005. Fontes alimentares de *Chagasia fajardi* (Lutz, 1904) (Diptera, Culicidae) de diferentes regiões do Brasil. Revista Brasileira de Zoociências 7: 181-184.

AYTEKIN, A. M.; TERZO, M.; RASMONT, P.; CAGATAY, N. 2007. Landmark based geometric morphometric analysis of wing shape in *Sibiricobombus* Vogt (Hymenoptera: Apidae: *Bombus* Latreille). Annales de la Société Entomologique de France 43: 95-102.

BAERG, D. C.; BOREHAM, M. M. 1974. Experimental rearing of *Chagasia bathana* (Dyar) using induced mating, and description of the egg stage. Journal of Medical Entomology 11: 631-632.

BERTI, J.; GUZMÁN, H.; LIRIA, J.; GONZÁLEZ, J.; ESTRA-DA, Y.; PÉREZ, E. 2011. Nuevos registros de mosquitos (Diptera Culicidae) para el estado Bolívar, Venezuela: Dos de ellos nuevos para el país. Boletín de Malariología Ambiental LI, (1): 59-69.

BOUYER, J.; RAVEL, S.; DUJARDIN, J.P.; DE MEEÜS, T.; VIAL, L.; THÉVENON, S.; GUERRINI, L.; SIDIBÉ, I.; SOLANO, P. 2007. Population structuring of *Glossina palpalis gambiensis* (Diptera: Glossinidae) according to landscape fragmentation in the Mouhoun river, Burkina Faso. Journal of Medical Entomology 44(5):788-95.

CALLE, D. A.; QUINONES, M. L.; ERAZO, H. F.; JARAMIL-LO, N. O. 2002. Morphometric Discrimination of females of five species of *Anopheles* of the subgenus *Nyssohrynchus* from Southern and Northwest Colombia. Memórias do Instituto Oswaldo Cruz 97: 1191-1195.

CAUSEY, O. R.; DEANE, L. M.; DEANE, M. P. 1945. Description of *Chagasia rozeboomi*, an anopheline from Ceará, Brasil. Journal of the National Malaria Society 4: 341-50.

DUJARDIN, J. P. 2003. Software for morphometric analysis PADwin. Available in: http://www.mpl.ird.fr/morphometrics. [Review date: 30 March 2011].

DUJARDIN, J. P. 2008. Morphometrics applied to medical entomology. Infection, Genetics and Evolution 8: 875-890.

^{*} Ta-III₅: the black band on the hindtarsus; Ta-III₅: the white band on the hindtarsus; Ta-III₄: the black band on the hindtarsus; Ta-III₃: the black band on the hindtarsus; Ta-III₄: the black band on the hindtarsus; Ta-III₃: the black band on the hindtarsus; Ta-III₄: the black band on the hindtarsus; Ta-III₃: the blac

Pop. 1: Bocaina Range National Park; Pop. 2: Corumbá Hydroelectric Plant, Goiás; Pop. 3: Itatiaia National Park; Pop. 4: Iguaçu National Park.

- DUJARDIN, J. P.; LE PONT, F. 2000. Morphometrics of a neotropical sandfly subspecies, *Lutzomyia carrerai thula*. C.R. Acad, Sci. Paris, Sciences de la vie / Life Sciences 323: 273-279. Hoboken. NJ. USA. 447 p.
- DUJARDIN, J. P.; SLICE, D. E. 2006. Contributions of morphometrics to medical entomology. Chapter 25 pp. 435-447. In: Tibayrenc, M. (Ed.). Encyclopedia of Infectious Diseases: Modern Methodologies. John Wiley & Sons, Inc.
- DUJARDIN, J. P.; BERMUDEZ, H.; SCHOFIELD, C. J. 1997. The use of morphometrics in entomological surveillance or sylvatic foci of *Triatoma infestans* in Bolivia. Disponible en http://www. mpl.ird.fr/morphometrics. html. [Review date: 11 September 2012].
- FORATTINI, O. P. 2002. Culicidologia Médica. 2nd Ed. Editora da Universidade de São Paulo. São Paulo. 864 p.
- GALINDO, P.; TRAPIDO, H. 1957. Forest mosquitoes associated with sylvan yellow fever in Nicaragua. The American Journal of Tropical Medicine and Hygiene 6: 145-52.
- HARBACH, R. E. 2007. The Culicidae (Diptera): a review of taxonomy, classification and phylogeny. In: Zhang, Z. Q.; Shear, W. A. (Eds.). Linnaeus Tercentenary. Progress in Invertebrate Taxonomy. Zootaxa 1668: 591-688.
- HARBACH, R. E.; HOWARD, T. M. 2009. Review of the genus Chagasia (Diptera: Culicidae: Anophelinae). Zootaxa 2210: 1-25.
- HARBACH, R. E.; KNIGHT, K. L. 1980. Taxonomists Glossary of Mosquito Anatomy. Plexus Publishing, Marlton, NJ, 415 p.
- JIRAKANJANAKIT, N.; LEEMINGSAWAT, S.; DUJARDIN, J. P. 2008. The geometry of the wing of *Aedes* (Stegomyia) *aegypti* in isofemale lines through successive generations. Infection, Genetics and Evolution 8: 414-421.
- KANOJIA, P. C.; PAINGANKAR, M. S.; PATIL, A. A.; GOKHALE, M. D.; DEOBAGKAR, D. N. 2010. Morphometric and allozyme variation in *Culex tritaeniorhynchus* mosquito populations from India. Journal of Insect Science 10: 138.
- KREUTZER, R. D. 1978. A mosquito with eight chromosomes: *Chagasia bathana* Dyar. Mosquito News 38: 554-558.

- LAURITO, M.; VISINTIN, A. M.; ALMIRÓN, W. R. 2008. *Culex saltanensis* morphological redescription of the immature and adult stages. Journal of the American Mosquito Control Association 24 (2): 203-10.
- MORAIS, S. A.; MORATORE, C.; SUESDEK, L.; MARRELLI, M. T. 2010. Genetic-morphometric variation in *Culex quinque-fasciatus* from Brazil and La Plata, Argentina. Memórias do Instituto Oswaldo Cruz 105 (5): 672-6.
- MORRISON, D. F. 1976. Multivariate Statistical Methods. Tokyo: McGraw-Hill Kogakusha Ltd.
- PY-DANIEL, L. H. R.; FERNANDES, C. C. 2005. Dimorfismo sexual em Siluriformes e Gymnotiformes (Ostariophysi) da Amazônia. Acta Amazonica 35: 97-110.
- REINERT, J. F. 2001. Revised list of abbreviations for genera and subgenera of Culicidae (Diptera) and notes on generic and subgeneric changes. Journal of the American Mosquito Control Association 17: 51-55.
- RUBIO-PALIS, Y. 1998. Caracterización morfométrica de poblaciones de *Anopheles (Nyssorhynchus) darlingi* del sur de Venezuela. Boletin de Entomologia Venezolana 13: 141-172.
- SILVA, J. S. 2011. Biodiversidade e Padrões alimentares de mosquitos (Diptera: Culicidae) em seis áreas brasileiras de preservação ambiental. Tese (Programa de Pós-graduação em Zoologia) Museu Nacional. Universidade Federal do Rio de Janeiro. Rio de Janeiro. 176 p.
- SOKAL, R. P.; SNEATH, P. H. 1963. Principles of numerical taxonomy. WH Freeman & Co. San Francisco and London. 359 p.
- SUMAN, D. S.; SHRIVASTAVA, A. R.; PARASHAR, B. D.; PANT, S. C.; AGRAWAL, O. P.; PRAKASH, S. 2009. Variation in morphology and morphometrics of eggs of *Culex quinquefasciatus* mosquitoes from different ecological regions of India. Journal of Vector Ecology 34 (2): 191-9.

Received: 8-Mar-2012 • Accepted: 9-Oct-2012